



## Characterization of the oxidative stress and inflammatory markers in metabolically healthy obese individuals

Hazhmat Ali

Department of Physiology and Pharmacology - College of Medicine - University of Duhok

### Article history

Received: 9 / 7 /2023

Revised: 3 / 9 /2023

Accepted: 10 / 9 /2023

DOI:

10.36320/ajb/v15.i3.13174

\*Corresponding Author:

Emails: [hazhmat.ali@uod.ac](mailto:hazhmat.ali@uod.ac)

**Abstract:** Obesity has emerged as a global health problem. Although various studies have linked obesity to a wide spectrum of diseases mainly diabetes and cardiovascular disease, little is known concerning involvement of oxidative stress and inflammation in non-diseased conditions. The current study aims to evaluate the levels of inflammatory and oxidative stress markers in sera of healthy obese individuals.

A case control study involved fifty healthy obese and fifty normal weight individuals. After obtaining relevant clinical data, sera were taken for further laboratory investigations including lipid profile, fasting serum glucose and HbA1c. Samples were also investigated for determining inflammatory and oxidative stress markers including highly sensitive C - reactive protein (hs-CRP), malondialdehyde (MDA) and gamma glutamyl transferase (GGT).

Total cholesterol, triglyceride and fasting blood glucose levels were significantly higher in healthy obese individuals compared to normal weight controls ( $p < 0.0001$ ,  $0.009$  and  $< 0.0001$  respectively). The hs-CRP, MDA and GGT were also statistically significantly higher in healthy obese ( $p < 0.001$ ,  $0.005$  and  $0.001$  respectively). Moreover, MDA was positively correlated with GGT and was statistically significant ( $p < 0.001$ ). The obtained findings suggest that, there may be an existence of a low grade chronic inflammation consistent with oxidative stress which might be considered as a risk factor for developing metabolic diseases and cardiovascular disturbances mainly diabetes mellitus, hypertension and stroke.

**Keywords:** obesity, inflammation, oxidative stress, body mass index

## 1.Introduction

Obesity has become one of the serious public health problems worldwide. It is defined as a massive accumulation of triacylglycerols in adipose tissues as a result of excessive energy intake compared to energy usage (1). Recent statistics indicate that the total number of population has exceeded 2 billion obese

constituting around 30% of the whole population globally. The Global Burden of Diseases in 2017 reported that the number of obese people have doubled in more than 70 countries since 1980 (2). This condition has a predominantly genetic basis and could probably be attributed to polygenic aspects. Other predisposing factors include higher energy intake and decreased physical activity

(3). While the physical consequences of obesity, such as arthritis, are debilitating and costly, the metabolic consequences mainly include insulin resistance, diabetes, fatty liver disease, coronary artery disease, hypertension and polycystic ovary syndrome (4).

Although numerous research have linked obesity to a wide spectrum of diseases (5), little is known concerning the role of inflammation and oxidative stress in obesity particularly in people who are metabolically healthy. Concerning inflammation, the traditional theory indicates that the basal systemic inflammation is gradually developed as people become older referring to the term inflammaging (6). The precise mechanism may be due to the contribution of inflammaging towards increasing risk of developing chronic diseases. However, this condition can be improved through non-pharmacological intervention strategies such as healthy diet consumption and physical activity performance (7). However, the current study will focus more on investigating the inflammation in young healthy obese individuals.

Oxidative stress is an imbalance between oxidative and anti-oxidative status in cells and tissues resulting in over production of free radicals and reactive oxygen species (ROS) (8). Excessive production of ROS molecules may attack cellular proteins and nucleic acids leading to cellular dysfunctions and subsequently loss of energy metabolism (9). It was reported that even high fat diet such as carbohydrates may promote generation of oxidative stress molecules represented by elevation in lipid peroxidation products coupled with a decrease in cellular anti-oxidant capacity which eventually increase the risk for developing chronic diseases (10).

Exploring oxidative stress and the inflammatory status in healthy obese will provide researchers with further insights towards better understanding of factors that predispose or protect obese individuals from metabolic disturbances. Moreover, will create ideas for therapeutic interventions that could

eventually improve the life quality of obese people. Taking this idea into consideration, the aim of the present study is to evaluate the role of inflammation and oxidative stress status in metabolically healthy obese as wells as assessment of glycemic and lipid status. Additionally, to find out the feasibility of any possible correlation between these markers and the metabolic status.

## **METHODS:**

### **Subjects:**

Following obtaining appropriate approval concerning research conduction ethics, the study was conducted. This case control study involved one hundred apparently healthy individuals; fifty obese with body mass index (BMI) > 30 kg/m<sup>2</sup> and fifty non-obese with BMI <25 kg/m<sup>2</sup>. Excluding criteria were hypertension, diabetes, smokers, pregnancy as well as people with history of cardiovascular disorders. A pre-requested questionnaire was filled in for each participant including basic demographic characteristics such as height, weight in addition to other necessary information. Arterial blood pressure was measured for study participants at resting condition using traditional mercury sphygmomanometer.

### **Specimen collection and storage:**

Five milliliters of venous blood was collected after ten hours of fasting in plain test tubes without anticoagulant. After coagulation, samples were centrifuged (at 1500 rpm for 5 min). The separated serum for each sample was divided into two aliquots. One is designated for the immediate assay of fasting glucose and lipid profile biomarkers. The other aliquot was stored at -20 C for subsequent assays such as highly sensitive C - reactive protein and other biomarkers.

### **Laboratory tests:**

Glycemic and lipid profile biomarkers including fasting serum glucose level, total

cholesterol (Tch), triglyceride (TG) and high density lipoprotein cholesterol (HDL) were assayed by automatic analyzer (Cobas 6000). Low density lipoprotein cholesterol (LDL) was calculated according to “Friedewald” equation (11).

Assessment of inflammatory markers included highly sensitive C-reactive protein (hs-CRP), assayed by enzyme linked immunosorbent assay “ELISA” using commercial kit (Monobind Inc., USA). The candidate markers for oxidative stress included malondialdehyde (MDA), a marker for lipid peroxidation as well as serum gamma glutamyl transferase (GGT). MDA was measured manually using thiobarbuturic acid method whereas GGT was measured using spectrophotometric method (Biolab, France).

### Statistical analysis:

Statistical analysis was performed using statistical package for social sciences (SPSS) Version 18 (Chicago, USA). All variables were expressed as mean  $\pm$  standard deviation. The clinical significance between study groups was determined using  $p$  value. The  $p$  values of 0.05 or less were considered statistically significant whereas the  $p$  values of 0.001 or less were considered statistically highly significant.

### RESULTS:

The current study involved a total of one hundred participants divided into two groups; the study group included fifty obese individuals whereas the control group represented 50 non obese. The study group had the mean age of 33 years vs. 31 years for controls. Both weight and body mass index (BMI) were statistically significantly higher in study group ( $p < 0.0001$ ) compared to controls. Both systolic and diastolic blood pressure didn't show any significant difference between study groups ( $p = 0.097$  and  $0.163$ ) respectively (table 1).

**Table 1: Demographic characteristics of study groups**

| Variables                            | obese<br>(no 50)  | non-<br>obese<br>(no 50) | $P$ value |
|--------------------------------------|-------------------|--------------------------|-----------|
| Age (years)                          | 33 $\pm$ 7.13     | 31 $\pm$ 6.25            | 0.139     |
| Height (cm)                          | 160 $\pm$ 5.04    | 162 $\pm$ 6.53           | 0.089     |
| Weight (kg)                          | 79.51 $\pm$ 12.14 | 64.2 $\pm$ 10.23         | < 0.0001  |
| Body mass index (kg/m <sup>2</sup> ) | 31.1 $\pm$ 4.27   | 24.5 $\pm$ 3.63          | < 0.0001  |
| Systolic blood pressure (mmHg)       | 118 $\pm$ 3.28    | 117 $\pm$ 3.88           | 0.097     |
| Diastolic blood pressure (mmHg)      | 76.7 $\pm$ 3.81   | 75.3 $\pm$ 4.22          | 0.163     |

Concerning lipid profile variables, both total cholesterol and triglyceride were statistically significantly higher in healthy obese compared to non-obese controls ( $p < 0.0001$  and  $0.009$ ) respectively. Other biomarkers including high density lipoprotein, low density lipoprotein and very high density lipoprotein showed no significant difference between study groups. The glycemic biomarker represented by serum blood glucose displayed a statistically significant elevation in study group compared to controls ( $p < 0.0001$ ), however, the hemoglobin A1c (HbA1c) was not statistically significantly different between study groups (table 2).

**Table 2: Lipid profile and glycemic biomarkers of study groups**

| Variables                            | obese<br>(no 50)   | non-<br>obese<br>(no 50) | $p$ value |
|--------------------------------------|--------------------|--------------------------|-----------|
| Total cholesterol (mg/dl)            | 183.51 $\pm$ 11.19 | 151.22 $\pm$ 14.8        | < 0.0001  |
| Triglyceride (mg/dl)                 | 100.62 $\pm$ 13.5  | 92.3 $\pm$ 11.9          | 0.009     |
| High density lipoprotein (mg/dl)     | 48.58 $\pm$ 7.06   | 47.12 $\pm$ 7.4          | 0.476     |
| Low density lipoprotein (mg/dl)      | 84.95 $\pm$ 4.51   | 84.65 $\pm$ 5.45         | 0.998     |
| Very low density lipoprotein (mg/dl) | 18.44 $\pm$ 1.9    | 18.48 $\pm$ 2.1          | 1.001     |
| Fasting blood glucose (mg/dl)        | 95.22 $\pm$ 8.14   | 78.39 $\pm$ 9.43         | < 0.0001  |
| HbA1c                                | 5.32 $\pm$ 0.43    | 5.25 $\pm$ 0.51          | 0.459     |

Both candidate biomarkers for oxidative stress; malondialdehyde and gamma GT were statistically significantly higher in healthy obese compared to non-obese controls ( $p = 0.005$  and  $0.001$  respectively). Moreover, highly sensitive C - reactive protein was also statistically significantly higher in study group ( $p < 0.001$ ) compared to controls (table 3).

**Table 3: Oxidative stress and inflammatory markers among study groups.**

| Variables  | obese<br>(no 50) | non-<br>obese<br>(no 50) | <i>P</i><br>value |
|--|------------------|--------------------------|-------------------|
| Malondialdehyde<br>(ng/ml)                         | 5.37 ±<br>0.98   | 2.11 ±<br>0.81           | 0.005             |
| Gamma GT (mg/dl)                                   | 32.77 ±<br>5.73  | 27.89 ±<br>6.11          | 0.001             |
| Highly sensitive c-<br>reactive protein<br>(mg/dl) | 6.44 ±<br>1.53   | 2.98 ±<br>1.09           | <<br>0.001        |

Pearson's correlation coefficient was used to find out the possible correlation between oxidative stress markers with the inflammatory hs-CRP. Interestingly, there was a statistically significantly positive correlation between MDA and GGT ( $r = 0.461$  and  $p < 0.001$ ) whereas the correlations between MDA – CRP and CRP – GGT were not significant (Table 4).

**Table 4: Pearson's correlation coefficient between inflammatory and oxidative stress biomarkers**

| Variables | <i>r</i> value | <i>p</i> value |
|-----------|----------------|----------------|
| MDA – GGT | 0.461          | < 0.001        |
| MDA – CRP | 0.189          | > 0.05         |
| CRP – GGT | 0.152          | > 0.05         |

Lastly, table 5 shows the correlation between BMI and inflammatory hs-CRP and oxidative stress markers. All markers showed a weak positive correlation with BMI except for GGT which displayed a moderate positive correlation and was statistically significant ( $r = 0.27$  and  $p < 0.05$ ).

**Table 5: Pearson's correlation coefficient between BMI with inflammatory and oxidative stress**

| Variables | <i>r</i> value | <i>p</i> value |
|-----------|----------------|----------------|
| BMI - MDA | 0.04           | 0.782          |
| BMI - GGT | 0.27           | < 0.05         |
| BMI - CRP | 0.05           | 0.730          |

## DISCUSSION:

Obesity has emerged as a global epidemic in the last decades. The reason behind that could be due to its obvious association with a variety of diseases mainly diabetes mellitus, cardiovascular diseases, nonalcoholic fatty liver disease (NAFLD), immune disorders and even cancer. This results in reduction in quality of life as well as shortened life span of people developing obesity (12, 13). Since it is apparent that extensive preclinical and clinical studies have confirmed the association between obesity with different categories of diseases, the aim of the present study was to determine the status of inflammation and oxidative stress in healthy obese individuals.

The demographic data confirmed the inclusion criteria of the study participants being apparently metabolically healthy individuals. The control group involved healthy people with normal body mass index to clearly identify the differences in biochemical and other laboratory parameters among study groups. Concerning the lipid profile status, obese populations had higher levels of serum total cholesterol and triglyceride compared to non-obese controls.

Since both groups are healthy subjects, this kind of elevations in lipid profile parameters could be attributed to dietary habits including excessive consumption of saturated fats and carbohydrates, smoking, alcohol intake and inadequate physical exercise (14, 15). Genetics may play a significant role in the development of elevated serum cholesterol levels such as familial hypercholesterolemia (FH) which is an autosomal dominant characterized by a substantial elevation in cholesterol and LDL levels from birth. It occurs as a result of genetic mutations in the LDL receptors or apolipoprotein B genes leading to a decrease in LDL metabolism (16, 17). Concerning the



glycemic status, the obese population displayed a higher levels in fasting blood glucose (except HbA1c) compared to normoglycemic controls. People with impaired fasting glucose are at a high risk for developing diabetes mellitus as well as cardiovascular diseases (18). Rhee et al. reported an association between obesity and glycemic status with developing heart failure in Korean population. It was concluded that people with elevated glucose levels and those with diabetes displayed an increased risk for developing heart failure compared to normoglycemic individuals (19).

C-reactive protein (CRP) is an acute phase protein that is considered a non-specific marker for inflammation. Its serum levels are elevated in all diseases involving activation of the inflammatory immune response mainly acute bacterial infections and autoimmune diseases (20). Our results indicated that the serum hs-CRP levels were substantially elevated in obese subjects compared to non-obese controls. The findings are consistent with previous studies showed that obesity is associated with elevated CRP levels. This association seems to be stronger in women specifically North American and Europeans (21). Another study revealed that the CRP levels were more elevated in depressive obese compared to non-depressives among German population (22). There are numerous studies that link CRP levels to obesity, decreased quality of life as well as cardiovascular disorders (23, 24). However, the only study found in the literature that investigated the CRP levels in healthy population was previously reported by McGill and Gronowski (25). It was concluded that even healthy obese develop increased levels of hs-CRP which was positively correlated with BMI.

Oxidative stress can result from imbalance between both pro-oxidant and antioxidant factors. To evaluate the oxidative stress status, two markers were used; malondialdehyde (MDA) and Gamma glutamyl transferase (GGT). The first one is widely used to assess the lipid peroxidation (a form of oxidative

stress) in biological samples whereas the later represents one of the essential liver enzymes. The current results emphasized that both markers were statistically significantly higher in the sera samples of obese individuals compared to control. These findings are in concordance with previous studies concluding that pronounced oxidative stress parameters such as myeloperoxidase activity (MPO) and nitric oxide levels were documented in obese people compared to controls (26). It was also reported that elevated MDA levels were positively associated with elevated diastolic hypertensive patients (27). Moreover, increased oxidative stress in accumulated fat was shown to be considered as an early initiator for metabolic syndrome (28). The current study revealed a positive correlation between MDA and GGT levels. This seems rational and supported by previous studies highlighting the validity of both parameters as early markers for oxidative stress and their association to cardiovascular diseases (29, 30).

After determining the role of inflammation and oxidative stress, the important question remains to be answered; what is the precise mechanism underlying these events. Moreover, what is the possible explanation behind the coexistence of these two essential players? The classical theory reveals that the accumulated adipose tissues respond to the highly diet intake by initiating an innate immune response. However, looking at the whole picture, the precise molecular mechanism may be explained by two possible points. Firstly, changes in the phenotype of the white adipose tissue (WAT) upon weight gain. This results in phenotype switch of WAT characterized by appearance of dysfunctional adipocytes initiating pro-inflammatory cytokine release and subsequently activating a low grade inflammatory immune response (31). Secondly, infiltration of the innate immune cells particularly macrophages into the already dysfunctional and inflamed adipocytes further exacerbates the condition. As a result, the inflamed adipose tissues become a reliable

source for secreting pro-inflammatory cytokines mainly interleukin 1 (IL-1), tumor necrosis alpha (TNF- $\alpha$ ) and interleukin 6 (IL-6) (32, 33). This low grade chronic inflammation results in altered metabolism that is also known as “metainflammation” (34).

Concerning the involvement of oxidative stress, the generated molecules results in cellular dysfunctions represented by altered cell signaling pathways, genetic mutations, impaired DNA damage eventually leading to reduction in biological activity, immune activation and inflammation (35, 36). Overall, long term existence of low grade chronic inflammation and oxidative stress in healthy obese can lead to initiation of pathologic milieu and subsequently increasing risk for developing chronic diseases.

## CONCLUSION:

The current study is considered one of the very few of its kinds documenting the presence of inflammation and oxidative stress in apparently healthy obese. The findings conclude that there may be an existence of a low grade chronic inflammation consistent with oxidative stress which might be considered as a risk factor for developing metabolic diseases and cardiovascular disturbances mainly diabetes mellitus, hypertension and stroke. Future studies should focus on investigating the molecular insights and detailed signaling pathways underlying these metabolic changes. Thorough understanding of the pathophysiology of these events may provide further insights towards developing therapeutic interventions.

## Conflicts of interest:

No conflicts of interest were reported.

## Funding:

The author declares that no funding was received from institutions or funding bodies for conducting this research.

## References

1. Mayoral LP, Andrade GM, Mayoral EP, Huerta TH, Canseco SP, Rodal Canales FJ, *et al.* (2020). Obesity subtypes, related biomarkers & heterogeneity. *Indian J Med Res.* Jan; 151(1):11-21.  
[https://doi.org/10.4103/ijmr.IJMR\\_1768\\_17](https://doi.org/10.4103/ijmr.IJMR_1768_17)
2. Afshin A, Forouzanfar MH, Reitsma MB, Sur P, Estep K, Lee A, Marczak L, Mokdad AH, *et al.* (2017). GBD 2015. Obesity Collaborators; . Health Effects of Overweight and Obesity in 195 Countries over 25 Years. *N Engl J Med.* Jul 6; 377(1):13-27.  
<https://doi.org/10.1056/NEJMoal614362>
3. Gregg EW and Shaw JE. (2017). Global Health Effects of Overweight and Obesity. *N Engl J Med.* Jul 6; 377(1):80-81.  
<https://doi.org/10.1056/NEJMe1706095>
4. Reilly JJ. (2017). Health Effects of Overweight and Obesity in 195 Countries. *N Engl J Med.* Oct 12; 377(15):1496.  
<https://doi.org/10.1056/NEJMc1710026>
5. Caballero B. (2019). Humans against Obesity: Who Will Win? *Adv Nutr.* Jan 1;10(suppl\_1):S4-S9.  
<https://doi.org/10.1093/advances/nmy055>
6. Dugan B, Conway J, Duggal NA. (2023). Inflammaging as a target for healthy ageing. *Age Ageing.* Feb 1; 52(2):afac328.  
<https://doi.org/10.1093/ageing/afac328>
7. Di Giosia P, Stamerra CA, Giorgini P, Jamialahamdi T, Butler AE, Sahebkar A. (2022). The role of nutrition in inflammaging. *Ageing Res Rev.* May; 77:101596.  
<https://doi.org/10.1016/j.arr.2022.101596>
8. Karam BS, Chavez-Moreno A, Koh W, Akar JG, Akar FG. (2017). Oxidative stress and inflammation as central mediators of atrial fibrillation in obesity and diabetes. *Cardiovasc*

- Diabetol. Sep 29;16(1):120. <https://doi.org/10.1186/s12933-017-0604-9>
9. Hu C, Yan Y, Ji F, Zhou H. (2021). Maternal Obesity Increases Oxidative Stress in Placenta and It Is Associated With Intestinal Microbiota. *Front Cell Infect Microbiol.* Aug 23; 11:671347. <https://doi.org/10.3389/fcimb.2021.671347>
  10. Rupérez AI, Gil A, Aguilera CM. (2014). Genetics of oxidative stress in obesity. *Int J Mol Sci.* Feb 20;15(2):3118-44. <https://doi.org/10.3390/ijms15023118>
  11. Naoto Fukuyama, Kazuhiro Homma, Noriaki Wakana, Kaori Kudo, *et al.* (2008). Validation of Friedewald Equation for Evaluation of Plasma LDL-Cholesterol. *J Clin Biochem Nutr.*; 43: 1–5.
  12. Rosen ED, Spiegelman BM. (2014). What we talk about when we talk about fat. *Cell.* Jan 16; 156(1-2):20-44. <https://doi.org/10.1016/j.cell.2013.12.012>
  13. Gissler MC, Anto-Michel N, Pennig J, Scherrer P, Li X, Marchini T, *et al.* (2021). Genetic Deficiency of TRAF5 Promotes Adipose Tissue Inflammation and Aggravates Diet-Induced Obesity in Mice. *Arterioscler Thromb Vasc Biol.* Oct; 41 (10):2563-2574. <https://doi.org/10.1161/ATVBAHA.121.316677>
  14. Riyami AA, Afifi MM. (2003). Prevalence and correlates of obesity and central obesity among Omani adults. *Saudi Med J.* Jun; 24(6):641-6.
  15. Karr S. (2017). Epidemiology and management of hyperlipidemia. *Am J Manag Care.* Jun; 23(9 Suppl):S139-S148.
  16. Vogt A. (2015). The genetics of familial hypercholesterolemia and emerging therapies. *Appl Clin Genet.*; 8:27-36. <https://doi.org/10.2147/TACG.S44315>
  17. Khera AV, Won HH, Peloso GM, *et al.* (2016). Diagnostic yield and clinical utility of sequencing familial hypercholesterolemia genes in patients with severe hypercholesterolemia. *J Am Coll Cardiol.*; 67(22):2578-2589. <https://doi.org/10.1016/j.jacc.2016.03.520>
  18. Neeland IJ, Poirier P, Després JP. (2018). Cardiovascular and Metabolic Heterogeneity of Obesity: Clinical Challenges and Implications for Management. *Circulation.* Mar 27; 137(13):1391-1406. <https://doi.org/10.1161/CIRCULATIONAHA.117.029617>
  19. Rhee EJ, Kwon H, Park SE, Han KD, Park YG, Kim YH, Lee WY. (2020). Associations among Obesity Degree, Glycemic Status, and Risk of Heart Failure in 9,720,220 Korean Adults. *Diabetes Metab J.* Aug; 44(4):592-601. <https://doi.org/10.4093/dmj.2019.0104>
  20. Norris T, Blodgett JM, Rogers NT, Hamer M, Pinto Pereira SM. (2022). Obesity in early adulthood and physical functioning in mid-life: Investigating the mediating role of C - reactive protein. *Brain Behav Immun.* May; 102:325-332. <https://doi.org/10.1016/j.bbi.2022.03.008>
  21. Choi J, Joseph L, Pilote L. (2013). Obesity and C-reactive protein in various populations: a systematic review and meta-analysis. *Obes Rev.* Mar; 14(3):232-44. <https://doi.org/10.1111/obr.12003>
  22. Chae WR, Nübel J, Baumert J, Gold SM, Otte C. (2022). Association of depression and obesity with C-reactive protein in Germany: A large nationally representative study. *Brain Behav Immun.* Jul; 103:223-231. <https://doi.org/10.1016/j.bbi.2022.04.024>

23. Han H, Cho YH, Lee SY, Park EJ, Kim YJ, Lee JG, et al. (2019). Elevated C-reactive protein level, obesity, and quality of life. *J Pak Med Assoc.* Dec; 69(12):1771-1776. <https://doi.org/10.5455/JPMA.298182>
24. Chandrasekhar J and Zaman S. (2020). Associations Between C-Reactive Protein, Obesity, Sex, and PCI Outcomes: The Fat of the Matter. *JACC Cardiovasc Interv.* Dec 28; 13(24):2893-2895. <https://doi.org/10.1016/j.jcin.2020.10.047>
25. McGill MR and Gronowski AM. (2018). Increased C - reactive protein in Healthy Controls. *Clin Chem.* Jan;64(1):242-243. <https://doi.org/10.1373/clinchem.2017.274746>
26. Jakubiak GK, Osadnik K, Lejawa M, Kasperczyk S, Osadnik T, Pawlas N. (2021). Oxidative Stress in Association with Metabolic Health and Obesity in Young Adults. *Oxid Med Cell Longev.* Jun 26; 2021:9987352. <https://doi.org/10.1155/2021/9987352>
27. Huang Y, Chen H, Liu Q, Hu J, Hu D, Huang Z, et al. (2023). Obesity difference on association blood malondialdehyde level and diastolic hypertension in the elderly population: a cross-sectional analysis. *Eur J Med Res.* Jan 24; 28(1):44. <https://doi.org/10.1186/s40001-022-00983-7>
28. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, Nakayama O, Makishima M, Matsuda M, Shimomura I. (2004). Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest.* Dec; 114(12):1752-61. <https://doi.org/10.1172/JCI21625>
29. de Oliveira Ulbrecht MO, Gonçalves DA, Zanoni LZG, do Nascimento VA. (2019). Association Between Selenium and Malondialdehyde as an Efficient Biomarker of Oxidative Stress in Infantile Cardiac Surgery. *Biol Trace Elem Res.* Jan; 187(1):74-79. <https://doi.org/10.1007/s12011-018-1378-y>
30. Shireen A. Ibrahim, Shelan H. Rasool, Hazhmat A. Ali. (2016). Serum gamma glutamyl transferase levels in women with polycystic ovary syndrome; relation to oxidative stress. *Duhok Medical Journal.*; (10): 1.
31. Pararasa C, Bailey CJ, Griffiths HR. (2015). Ageing, adipose tissue, fatty acids and inflammation. *Biogerontology.* Apr; 16(2):235-48. <https://doi.org/10.1007/s10522-014-9536-x>
32. Kawai T, Autieri MV, Scalia R. (2021). Adipose tissue inflammation and metabolic dysfunction in obesity. *Am J Physiol Cell Physiol.* Mar 1; 320(3):C375-C391. <https://doi.org/10.1152/ajpcell.00379.2020>
33. Scalia R. (2013). The microcirculation in adipose tissue inflammation. *Rev Endocr Metab Disord.* Mar;14(1):69-76. <https://doi.org/10.1007/s11154-013-9236-x>
34. Harman-Boehm I, Blüher M, Redel H, Sion-Vardy N, Ovadia S, Avinoach E, Shai I, et al. (2007). Macrophage infiltration into omental versus subcutaneous fat across different populations: effect of regional adiposity and the comorbidities of obesity. *J Clin Endocrinol Metab.* Jun; 92(6):2240-7. <https://doi.org/10.1210/jc.2006-1811>
35. Rani V, Deep G, Singh RK, Palle K, Yadav UC. (2016). Oxidative stress and metabolic disorders: Pathogenesis and therapeutic strategies. *Life Sci.* Mar 1; 148:183-93. <https://doi.org/10.1016/j.lfs.2016.02.002>
36. Raut SK and Khullar M. (2023). Oxidative stress in metabolic diseases: current scenario and therapeutic relevance. *Mol Cell Biochem.* Jan; 478(1):185-196. <https://doi.org/10.1007/s11010-022-04496-z>