



Molecular technique for identification and characterization of *Acinetobacterbaumannii* *

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Abstract

The present study involved isolation and characterization of *Acinetobacter baumannii* from patients suffering from urinary tract infection (UTI), wound infection and vaginitis . A total of 200 clinical samples were collected from patients admitted to Al-Zahraa hospital, Al- Hakeem hospital and Al – Sadder teaching city from September 2011 to January 2012. Identification of isolates has been carried out depending on biochemical test and polymerase chain reaction technique (PCR) . Our study proved that PCR technique depending on *rpoB* gene was a specific method for identification of *A.baumannii* compared with *gyrB* gene in which 26 isolates have been identified which included 13 (36%) isolates from UTI, 7 (19%) isolates from wounds infection and 5 (13%) isolates from vaginitis.

Key word : (*Acinetobacter baumannii* , urinary tract infection , PCR , *gyrB* gene , wounds infection).

Introduction:

Acinetobacter is a gram negative coccobacilli that is strictly anaerobic, catalase positive and oxidase negative. It is widely distributed in nature, being found in soil and water. The taxonomy of *Acinetobacter* revealed at least 32 named and unnamed species that have been described [1], of those *A.baumannii* and closely related unnamed genomic species 3 and 13TU which were the most clinically relevant. These three species are commonly grouped together with the environment species *A.calcoaceticus* in the *A.calcoaceticus* – *A.baumannii* complex because of the difficulty in separating these species by using phenotypic method [9; 11].

Manual and semi – automated commercial identification system such as API 20NE, and VITEK2 don't differentiate among these species , so that gene sequences including *gyrB*, *recA* gene have been proposed to delineate species within the genus *Acinetobacter* [2; 3; 4].The use of protein-encoding genes as phylogenetic markers is now a common approach[4; 5; 6]. Many investigations have demonstrated that sequences from protein-encoding genes can accurately predict genome relatedness and may replace DNA–DNA hybridization for species identification and delineation in the future [7; 8].The use of *gyrB* as a means to identify *Acinetobacter* isolates to the species level is not new, it was demonstrated over a decade ago that nucleotide and amino-acid sequences can both be used for taxonomic purposes, and that they correlate with DNA–DNA hybridization[8].The rate of molecular evolution inferred from *gyrB* gene sequences is faster than that inferred from16S rRNA gene sequences [3]. Several molecular studies which depend on amplification of rDNA restriction

analysis is recognized as providing good discrimination, but can be time-consuming, while tRNA fingerprinting does not discriminate between *A. baumannii* and genomic sp. 13TU.

Sequencing of the *rpoB* gene and its flanking spacer regions, and of the 16S, 23S rRNA gene spacer region, have been proposed for identification of *Acinetobacter* isolates to the species level [11; 12]. Various protein-encoding genes such as *recA*, *groEL*, *rpoB*, *rpoD* and *gyrB* have been used for the classification of bacteria at the intragenetic level [2;4]. To focus the light on an important pathogenic bacteria this study was carried out. to explain a perfect method in identification and diagnosis of *A. baumannii*.

Materials and Methods:

* Collection of samples:

A total of 200 samples were recorded from patients suffering from burn infection (50 sample), vaginal infection (50 sample) and urinary tract infection (100 sample) who admitted to AL-Zahar Teaching hospital, AL-Hakem hospital and AL-Sader Teaching city, during a period from September 2011 to January 2012. All samples were collected in a proper way to avoid any possible contamination.

* Laboratory diagnosis:

Primary identification and characterization of *A. baumannii* from other isolates were performed according to depending on the morphological properties of colonies growing on blood agar base after incubation at 44°C for 24 hr. These properties involved: colony shape, size, color, translucency, by biochemical test which include: oxidase, catalase, Indole MR-VP, simon citrate, KIA and hemolysin tests. *Molecular identification by PCR.

DNA extracted from bacterial cell by alkaline lysis method described by Sambrook and Russell (2001) was used as a template in a specific PCR for diagnosis of *A. baumannii* by *gyrB* and *rpoB* gene. A set of primer (table 1) was used for amplification of *gyrB* and *rpoB* gene using reaction mixture (table 2) with final volume of 25 µl for *gyrB* and 50 µl for *rpoB* gene.

Table (1): Primers of gene amplification

Primer	Sequence	PCR products
Sp4F	CACGCCGTAAGAGTGCATTA	294 bp 490 bp
Sp4R	AACGGAGCTTGTCAGGGTTA	
Sp2F	GTTCTGATCCGAAATTCTCG	
Ac696F	ATAYCGYAAAGAYTTGAAAGAAG	397 bp
Ac1093R	CMAACCCYTTGTTMCCRTGA	

Table(2): Volume of reaction in a PCR mixture for *gyrB*, *rpoB* gene

Component	<i>gyr B</i> gene	<i>rpoB</i> gene
	Volume (μl)	Volume (μl)
Go Tag Green Master Mix,	12.5	25
Upstream primer	5*	1.0
Downstream primer	2.5	1.0
DNA template	5	2.5
Nuclease – free water	—	18
DMSO	—	2.5
Total	25	50

PCR amplification was performed in a PCR thermo cycler (Biometra ,USA) using the program explained in table (3). PCR products and 100 bp DNA ladder (Promega, USA) were visualized by 2 % agarose gel electrophoresis and stained with ethidium bromide. The electrophoresis was performed under 60 Volts for 80 minutes .UV-transilluminator was used to visualize DNA band and gel was photographed using digital camera [7].

Table (3): PCR condition for amplification of *gyrB* and *rpoB* gene

Primer	No. of cycle	PCR condition		
		denaturation	Annealing	Extension
*Sp4F Sp4R Sp2F	25	94°C 2 min	94°C for 1 min 60°C for 30 S 72 °C for 1 min	72 °C for 10 min
**Ac696F Ac1093R	35	94°C 5 min	94°C for 1 min 52 °C for 1 min 72 °C for 1 min	72 °C for 10 min

*Primers of *gyrB* gene , **Primers of *rpoB* gene.

Result and discussion:

A total of 200 samples were collected from patients suffering from Urinary tract infections (100) , vaginal infections (50) , and wound infections (50) . All samples were cultured on blood agar base under 44 °C for 24 hrs. A biochemical test and

molecular method by using specific gene *gyrB* and *rpoB* gene were used for the identification of *Acinetobacter* spp. According to the biochemical test 100 isolates were suspected to be *A.baumannii*. The results of amplification by PCR for *gyrB* gene were shown in figure(1). *A.baumannii* was identified when two amplicons were described with molecular weight (294) bp and (490) bp, while *A. genomic*s was identified when one amplicon with (294) bp was observed. Our results showed that 36 (36%) isolates were identified as *A.baumannii* while 15 (15%) isolates were identified as *A. genomic*s, other isolates show no PCR product

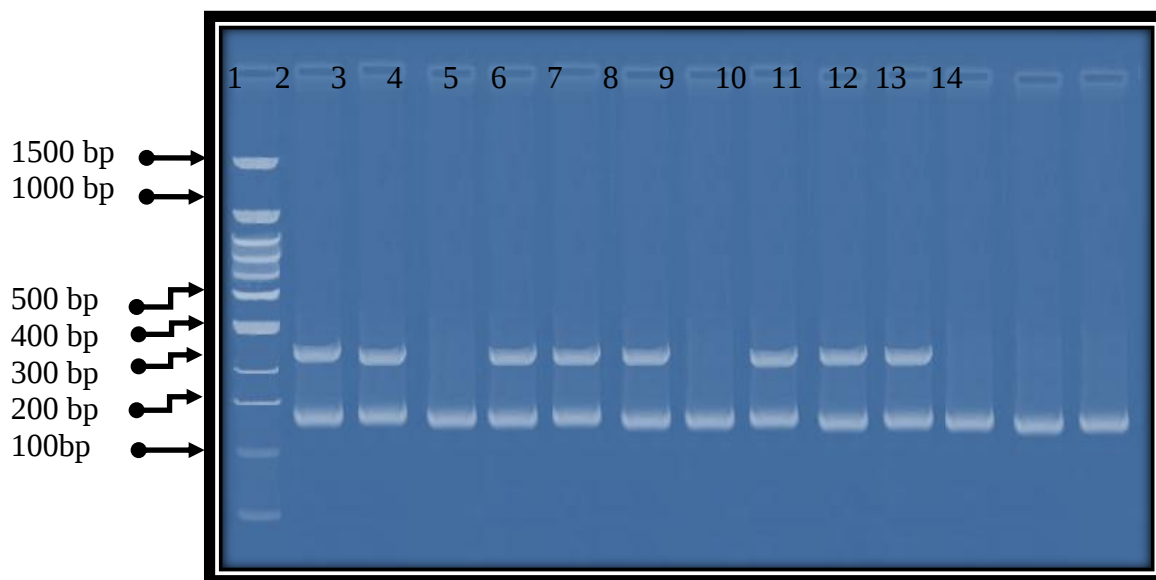


Figure (1): Identification of *A.baumannii* by amplification of *gyr B* gene by PCR (amplicons size 294 bp and 490 bp). Line 1: Ladder DNA (100bp). Line 2,3,5,6,7,9,10,11: *A. baumannii* Line 4,8,12,13,14: *A. genomic*s .

On the other hand, Figure (2) showed the results of amplification of *rpoB* gene ,it explained that out of 36 isolates which were identified by *gyrB* gene, only 26 (72%) isolates were identified as *A. baumannii* with PCR product (397) bp

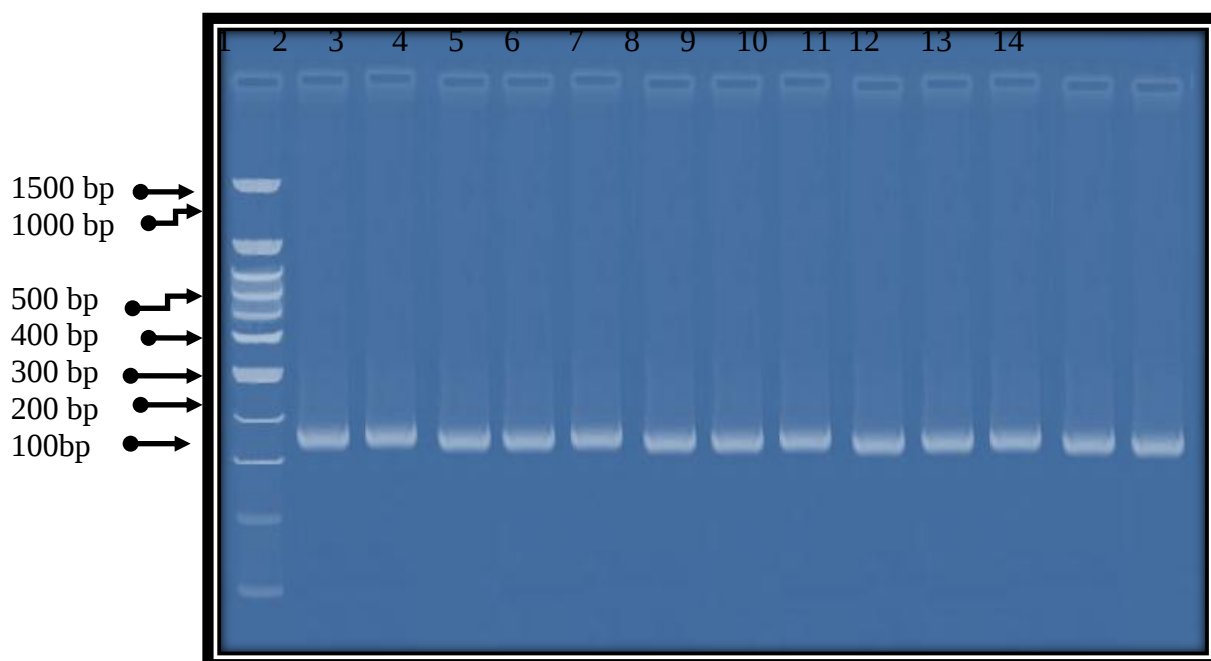


Figure (2): Identification of *A. baumannii* by amplification of *rpoB* gene by PCR (amplicon size 397 bp).

Line 1: Ladder DNA (100bp).Line 2 - 14: *A.baumannii* isolates.

Because of increasing pathogenic importance of *Acinetobacter spp*, a development of reliable identification methods for these strain was required. Many identification schemes depending on DNA homology, phenotypic characters , and compression of cell envelope and outer membrane protein pattern yield in consistent result [9 ; 13 ; 14 ; 15].

Phenotypically many geno-species are very similar and are not readily distinguishable [8].The currently used 16S r RNA gene sequencing determination have failed to distinguished closely related genomic species of *Acinetobacter* due to its extremely low polymorphic nature [9; 13].

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* تقنيات جزيئية لتشخيص وتوصيف بكتيريا *Acinetobacter baumannii*

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الخلاصة

تضمنت الدراسة الحالية عزل وتشخيص بكتيريا *Acinetobacter baumannii* من إخماج المجاري البولية ، الحروق، والمهبل . جُمعت 200 عينة سريرية من المرضى المراجعين لمستشفى الزهراء والحكيم ومدينة الصدر التعليمية للفترة من سنة 2011 ولغاية 2012 . وتم الاعتماد على الاختبارات البايوكيميائية وتقنية تفاعل السلسلة المتبلمر PCR في تشخيص العزلات وقد اثبتت الدراسة الحالية ان افضل طريقة في التشخيص هي تقنية ال-PCR باستخدام الجين التشخيصي *rpoB* اذ تم عزل وتشخيص 26 عزلة وبواقع (7) من إخماج الجروح. (13) عزلة من إخماج الادرار و (5) عزلات من إخماج المهبل.