

Role of serotonin on pituitary-thyroid functions and antioxidants in diabetic male rats

Fouad Ziedan Hamzah¹

Sumayah Faruq Kasim²

¹Faculty of Veterinary Medicine, University of Kufa, Iraq. ²College of Health and Medical Techniques, Middle Technical University, Iraq.

Corresponding author: SFK, e-mail: <u>sumayah.faruq@mtu.edu.iq</u>; ORCID ID: <u>0000-0003-1809-6832</u> Co-author: FZH, e-mail: <u>fouadz.alquraishi@uokufa.edu.iq</u>; ORCID ID: <u>0000-0003-0642-3381</u>

Received date: 12 July. 2023 Accepted: 21 Sep. 2023 page: (24-32) Published: 30 Dec. 2023

Abstract

Diabetes mellitus causes an increase in oxidative stress. Serotonin is a hormone its level in circulation reduced in diabetes; recently, it has been found to possess antioxidant qualities. The purpose of the study was to look at the impact of diabetes on pituitary-thyroid functioning, and serotonin role in modulating these effects in a streptozotocin-induced diabetes mellitus rat model. The thirty-tow healthy male rats with a body weight of 180-200 gm were split equally into 4 groups, group [1]: control, group [2]: Serotonin, group [3]: diabetic, and group [4]: Serotonin-diabetic. The rats were weighed and serum levels of glucose, insulin, malondialdehyde, superoxide dismutase, catalase, glutathione peroxidase, serotonin, thyroid stimulating hormone, thyroxine, and triiodothyronine were measured. Our results presented that there was a significant increase in T3, T4, glucose, and MDA in the diabetic group compared to the control, while a significant decrease in T3, and T4, glucose, and MDA in the Serotonin with diabetic group compared to diabetic group. Furthermore, a significant decrease in TSH, serotonin, insulin and GPX, CAT, and SOD antioxidants in the diabetic group compared to control, while a significant increase in serum TSH, serotonin, insulin and CAT, GPX and SOD antioxidants in the Serotonin+diabetic group compared to diabetic group as well as a significant increase in serum insulin in the Serotonin group compared to all other groups. Serotonin plays a protective role against diabetes-induced and pituitary-thyroid disorder caused by diabetes which might be because of its antioxidant qualities and ability to maintain glucose and insulin balance.

Keywords: Anti-oxidants, Diabetes Mellitus, Oxidative stress, Pituitary-thyroid, Serotonin.

Copyright: © 2023 Hamzah & Kasim. This is an open access article distributed under the terms of the <u>Creative Commons</u> <u>Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

INTRODUCTION

Several metabolic conditions are included in diabetes mellitus identified by hyperglycemia induced by insulin production, activity, or both. Diabetes damages, dysfunctions, and fails various organs over time, including the kidneys, nerves, eyes, heart, and blood vessels [1, 2]. It is common practice to create a rat model of type I diabetes with a single treatment of streptozotocin (STZ), which selectively kills beta cells in pancreatic islets that produce insulin [3].

Different functions of 5hydroxytryptamine are found in the nonneuronal and neuronal mechanisms. It functions like a neurotransmitter. mitogen, and a hormone. Serotonin was given this name just after Latin word serum as well as the Greek word tonic since it was to be a vessel-constrictor assumed contained in platelets at the period of its discovery (1918) [4, 5]. The physiological consequences of 5-hydroxytryptamine are initiated through its absorption inside the cells by 5-hydroxytryptamine transporter (5-HT) and successfully completed by membrane-bound 5-hydroxytryptamine transporter receptors [6].

Serotonin is a biologically active monoamine neurotransmitter in which is produced centrally inside neurons from L tryptophan in considerable quantities in neurons of the central nervous system (CNS) and peripherally in platelet granules in the gastrointestinal tract (GIT) [7]. Serotonin regulates a variety of vascular dynamics, including vascular resistance, blood pressure management, and platelet aggregation [8]. Seven distinct types of serotonin receptors (5-HT1 to 5-HT7), which are dispersed throughout the body, and mediate its activity [9].

Following a 5-hydroxytryptophan preload, a precursor of serotonin, it has long been recognized that 5-HT and insulin colocalize in the same vesicle in pancreatic β cells. Additionally, serotonin has been connected to the control of glucose levels [10, 11]. 5-HT has recently been discovered to be a placental lactogen's downstream molecule, regulating how β -cells adapt to pregnancy [12]. Tph1 expression in pancreatic β -cells increases dramatically as a result of lactogens released throughout the pregnancy, which causes a large synthesis of serotonin in β -cells [12]. Through the 5-HT3 receptor, islet serotonin improves beta cell glucose responsiveness, leading to greater islet glucose-stimulated insulin secretion (GSIS) [13, 14]. Additionally, mid-gestation is when the expression of the 5-HT2B receptor rises to encourage β-cell

proliferation and increase β -cell mass, whereas the 5-HT 1D receptor expression rises near the end of a pregnancy to decrease β -cell mass [12].

In dietary insulin resistance, 5-HT regulates insulin secretion as well. In response to an HFD, glucose intolerance was observed in -cell-specific Tryptophan hydroxylase 1 gene (Tph1) Knock-Out (KO)and 5-Hydroxytryptamine 3A(Htr3a) Knock-Out (KO) Receptor mice, but not in a wild-type genotype (WT) mice. The Glucose-stimulated insulin secretion(GSIS) was disrupted in islets extracted from high-fat diet (HFD-fed) Htr3a KO and -cell-specific Tph1 KO mice, and 5-HT administration increased insulin production from the -cells of the latter animals but not the Htr3a KO islets [13]. Additionally, the embryonic pancreas has been found to produce 5-HT. Particularly, at the time just after birth, when β -cells are actively multiplying, Tryptophan hydroxylase 1 geneexpression is increased in newborn β --cells [15]. Since 5-HT is produced by -cells during the perinatal and during pregnancy, period two physiological conditions that promote -cell proliferation, it is thought that 5-HT is a critical controller of -cell proliferation and insulin release [12-14, 16].

The purpose of this research was to look at diabetes mellitus' effects on the pituitary-thyroid functioning and the function of serotonin in modulating these results in a streptozotocin-induced diabetes mellitus rat model.

MATERIALS AND METHODS Animal Ethical Approval:

This study was approved by the ethics committee of the Faculty of Veterinary Medicine, University of Kufa and conforms to the Guide for the Care and Use of Laboratory Animals (No.9129).

Sampling:

Thirty-two healthy albino male adult rats weighing 180–200 g were used in this investigation; they were bought from the animal house at the Faculty of Veterinary Medicine at the University of

2023

Kufa. Rats were housed at physiology animal house at Faculty of Medicine -University of Kufa in four-per-cage steel wire cages under sanitary conditions [17-19]. Before the trial began, rats spent one used their week getting to new surroundings. They were kept at ambient temperature, a 12-hour light/dark cycle maintained, and unrestrained access to water. Rats were acclimated to their new surroundings for one week prior to the experiment.

Rats were split into four equal groups at random. In group 1, the control each rat received a single group, intraperitoneal dose of 0.5 mL of saline for 10 days. In group 2, each rat received a single intraperitoneal dose of freshly synthesized serotonin hydrochloride [Sigma Aldrich Co., USA powder] at a quantity of 1.6 mg/kg of body weight (b.w) dissolved in saline for 10 days [20], group 3: diabetic group, which was given an intraperitoneal injections of fresh manufactured streptozotocin[Sigma Aldrich Company, USA] at a dose of 60 mg/kg diluted in saline to develop experimental diabetes [21] and overnight the fasted rats, The fasting glucose level was measured using a glucometer after 72 hours [ACCUCHEK, Rhoche Diagnostics, Germany], this study included rats with blood glucose levels more than 360 mg/dl. [22], group [4]: Serotonin-treated diabetic rats received a single 1.6 mg/kg intraperitoneal injection of serotonin of [b.w] daily for 10 days.

Biochemical Tests:

Serum levels of glucose: So according Tietz, (1995) [23], it is tested using a 546 nm spectrophotometer (Spectronic 3000 Array, Germany) and glucose enzymatic (GOD-PAP) - liquizyme Kits from Biotechnology, Egypt. Serum levels of insulin: By utilizing Kits for Enzyme Amplified Sensitivity Immunoassay (KAP1251-INS-EASIA), as according to Temple et al., (1992)[24] from (BioSource Europe S.ABelgium).

Triiodothyronine (T3), (T4) and TSH Total ELISA Kit were commercially supplied by Elabscience, Serotonin was carried out by using commercially available kits supplied by Immusmol SAS.

Serum antioxidant system evaluation: Measurements of Superoxide Dismutase (SOD), MDA, GlutathionePeroxidase (GPX) catalase (CAT): Elabascience Biotechnology Inc.'s ELISA kit from China is used to evaluate the serum level of Superoxide Dismutase (SOD).

Statistical analysis:

The data underwent an analysis of variance, and the significant differences at $P \le 0.05$ were assessed by ANOVA, one-way by utilizing the statistical software's sigma statistical.

RESULTS

Effect of Serotonin on T3, T4, TSH and Serotonin levels showed in Table (1) which demonstrates that, in comparison to the control group, the diabetic group experienced a significant ($P \le 0.05$) increase in serum T3 and T4, whereas the Sero with diabetic group experienced a significant (P≤0.05) decrease in serum T3 and T4 in comparison to the diabetic group. However, there was a substantial (P<0.05) drop in serum TSH and serotonin in the diabetic group when compared to the control group and a significant ($P \le 0.05$) rise in serum TSH and serotonin in the Sero with diabetic group when compared to the diabetic group.

Parameters	Т3	T4	TSH	Sero
Groups	ng/ml	µlU/ml	µlU/ml	ng/ml
Control	2.058±0.027 b	7.104±0.048 d	1.078±0.100 b	7.106±0.045 b
Sero+diabetic	2.084±0.042 b	8.074±0.089 c	1.260±0.426 b	9.638±0.316 a
Sero	1.516±0.219 c	9.072±0.050 b	1.934±0.449 a	9.290±0.430 a
diabetic 4.052±0.04		10.316±0.071 a	0.500±0.141 c	4.130±0.136 c
LSD	0.165	0.094	0.430	0.369

Table (1): Effect of Serotonin on T3, T4, TSH and Serotonin levels in diabetic adult male r	ats
---	-----

Significant group differences are indicated by tiny letters at $[p \le 0.05]$ level.

Effect of Serotonin on Glucose and Insulin levels presented inTable (2) which demonstrates that, the diabetic group had a substantial (P \leq 0.05) rise in glucose compared to the control group, whereas the Sero with diabetic group had a significant (P \leq 0.05) drop in glucose compared to the diabetic group. While there was a significant (P \leq 0.05) decrease in serum insulin in the diabetic group when compared to the control group, there was also a significant (P \leq 0.05) increase in serum insulin in the Sero with diabetic group when compared to the diabetic group, as well as a significant (P \leq 0.05) increase in serum insulin in the Sero group when compared to all other groups.

Parameters	Glucose	Insulin	
Groups	mg/dl	µIU/ml	
Control	85.188±1.959 c	27.540±0.359 b	
Sero+diabetic	165.538±0.802 b	18.52±0.297 c	
Sero	80.126±0.026 d	29.322±0.393 a	
diabetic	579.656±4.855 a	8.116±0.063 d	
LSD 3.501		0.420	
a: : a 1:00			

Significant group differences are indicated by tiny letters at $[p \le 0.05]$ level.

Finally, the effect of Serotonin on CAT, GPX, SOD antioxidants and MDA levels illustrated in Table (3) which demonstrates that MDA significantly increased in the diabetic group (P \leq 0.05) when compared to the control group, but significantly decreased in the Sero with diabetic group (P \leq 0.05) when compared to the diabetic group and the

control group. On the other hand, the diabetic group had а substantial $(P \le 0.05)$ drop in CAT, GPX, and SOD antioxidants compared to the control group, whereas the Sero with diabetic group had a significant $(P \le 0.05)$ increase in all antioxidants GPX, CAT and SOD compared to the diabetic group.

Table (3): Effect of Serotonin on CAT, GPX, SOD antioxidants and MDA levels in diabetic
adult male rats

Parameters	CAT	GPX	MDA	SOD
Groups	U/ml	µmol/L	nmol/L	ng/Ml
Control	758.25±26.925 c	91.202±1.170 b	4.108±0.134 c	7.036±0.228 a
Sero+diabetic	911.61±5.090 a	92.148±1.287 b	5.538±0.360 b	6.026±0.392 b
Sero	876.04±13.627 b	102.448±0.979 a	3.136±0.136 d	7.152±0.229 a
diabetic	349.32±23.534 d	65.584±0.852 c	9.058±0.165 a	3.428±0.451 c
LSD	26.963	1.598	0.322	0.471

Significant group differences are indicated by tiny letters at $[p \le 0.05]$ level.

DISCUSSION

The most common metabolic illness, people diabetes, affects from all socioeconomic backgrounds. Reduced insulin release from the pancreatic β -cells or diminished insulin activity on its target organs may both contribute to the comprehensive condition. Therefore. knowledge of the control of insulin secretion is necessary to comprehend how to prevent and cure diabetes. Serotonin is a hormone that we detect in our study as an improver to modulate the release of insulin which in turn regulates the levels of glucose in the blood. To have a better knowledge of type 1 and type 2 diabetes mellitus causes and the young-onset maturity of diabetes, several animal models have been developed [25–27], Our findings showed a significant increase in serum glucose levels in the STZ-diabetic group as compared to the control group, while a significant decrease in serum glucose levels in the Sero with diabetic group as compared to STZ-diabetic group, However, a significant decrease in serum insulin was observed in the STZdiabetic group when compared to the control group, while significant increases in serum insulin were observed in the Sero with diabetic group when compared to the STZ-diabetic group as well as in the Sero group when compared to all other groups. These findings obviously show the ameliorative role of serotonin hormone on insulin and glucose levels in diabetic rats. As described in the introduction part, mounting evidence suggests that 5-HT has a beneficial impact on the body's overall glucose homeostasis. However, many regulatory features of insulin secretion are unclear because of the intricate machinery involved. The function of serotonin in β cells is one topic that has baffled scientists for more than three decades [28, 29]. Within β -cells, 5-HT is synthesized [30], it in their secretory is kept βgranules alongside with insulin [11], and when pancreatic islets are activated, it is also co-secreted with glucose [31, 32].

Prior research has shown a unique mode of serotonin activity in blood platelets that is dependent on the hormone's longterm covalent attachment to signaling "serotonylation." The proteins, or researchers found similar process in pancreatic beta cells as well. Serotonylation controls the release of storage granules from these cells, just like it does in thrombocytes. Serotonin regulates insulin secretion, which is the most significant hormone in controlling blood glucose levels in both humans and animals, under normal circumstances. In addition to the study's significance to the knowledge of serotonin's function in the common illness diabetes. the unique example of serotonylation serves as an illustrative illustration physiological of the significance of protein monoaminylation in general. Similar effects can be achieved by other monoaminergic hormones such as histamine, dopamine, and norepinephrine. Monoaminylation, like protein phosphorylation, has a significant influence on a number of biological processes, and Berlin-based researchers have embraced challenge to identify the them. The findings. which were reported by researchers from Berlin, provide an explanation for what serotonin is doing in the pancreatic beta cells and broaden our comprehension of the function of protein physiological monoaminvlation in processes that are important for illness [33].

In this study we focused our interest in the relation between pituitary-thyroid function, antioxidants and serotonin; this is due to the lack of previous studies on this aspect. Furthermore, our data presented that as compared to the control group, the STZdiabetic group showed a substantial increase in blood T3, T4, and MDA and a significant decrease in serum TSH. serotonin, and CAT, GPX, and SOD antioxidants. The Sero with diabetic group showed a significant increase in serum T3, T4, and MDA, as well as a significant increase in serum TSH, serotonin, and CAT, GPX, and SOD antioxidants when

2023

compared to the STZ-diabetic group. Several scientists and researchers have found that there is a dual relationship between the thyroid gland and diabetes, a disease caused by insulin resistance in the body, and insulin is a hormone responsible for controlling blood sugar, it proven a direct relationship between hypothyroidism and diabetes and vice versa, high blood sugar by increasing the amount of sugar secreted by the liver and/or increasing the body's resistance to insulin can lead to high levels of thyroid hormones. Despite decades of research, much is still unknown about serotonin, despite the fact that it affects many aspects of health and has a wide range of effects. Apparently, thyrocytes [the cells that make up thyroid] can be stimulated into action by serotonin, as stated by Dobberstein [34], it stimulates thyroid cell proliferation and apoptosis, this suppose aligned with our findings in improving T3 and T4 levels as a result of the serotonin effect in ameliorating the oxidative stress resulting from diabetes on thyroid cells.

Numerous physiopathological conditions and degenerative illnesses are linked to the oxidation of biological components including DNA, proteins, or lipids [35-38].

These research results taken together compelled us to describe the ways in which 5-HT exerts its antioxidant capabilities. Numerous experimental investigations have demonstrated the important antioxidant effects of 5-HT. The molecule's chemical system has produced strong free radical scavenging characteristics in vitro [39-42]. Additionally, 5-HT has been utilized to stop the production of free radicals caused by chemicals in vitro [43].

We assumpted that the 5-HT has a modulating effects on T3, T4, CAT, GPX and SOD antioxidants, results clearly show 5-HT efficiency, suggesting its direct involvement in modulating the effects of diabetes mellitus in rats model induced by streptozotocin as well.

CONCLUSION

Serotonin may have a preventive role against diabetes-induced pituitary-thyroid disturbances because of its antioxidant characteristics and maintaining insulin and glucose concentrations.

ACKNOWLEDGEMENT

The authors want to express their thanks and recognize Prof.Dr. Ali Faruq Kasim's invaluable assistance in conducting the statistical analysis.

CONFLICT OF INTEREST

There is no conflict of interest.

REFERENCES

- 1. Cho NH, Shaw JE, Karuranga S, Huang Y, da Rocha Fernandes JD, Ohlrogge AW. Malanda B. IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and for Diabetes projections 2045. Research Clinical and Practice.2018;138:271-281. DOI:10.1016/j.diabres.2018.02.0 23
- Ugbaja RN, FatokunTP, Akinloye DI, James AS, OnabanjoOO, Akinloye OA. Propolis ethanol extract abrogates hyperglycemia, lipotoxicity, and lowered hepatic poly [ADP-ribose] polymerase protein level in male albino rats. Journal of Diabetes & Metabolic Disorders. 2021;20[1]:683-696. DOI:10.1007/s40200-021-00800-8
- 3. Rerup CC. Drugs producing diabetes through damage of the insulin secreting cells. Pharmacological reviews.1970;22:485-518.
- 4. Janeway TC, Richardson HB, Park EA. Experiments on the vasoconstrictor action of blood serum Archives of Internal Medicine. 1918;21[5]:565-603. DOI:10.1001/archinte.1918.0009 0100002001
- 5. Zucker MB. A study of the substances in blood serum and platelets which stimulate smooth muscle. American

Vol. (14) No. (2)

Journal of Physiology.1944;142:12-26.

DOI:10.1152/ajplegacy.1944.142.1.12

- Wade PR, Chen J, Jaffe B, Kassem IS, Blakely RD, Gershon MD. Localization and function of a 5-HT transporter in crypt epithelia of the gastrointestinal tract. The Journal of neuroscience: the official journal of the Society for Neuroscience.1996;16:2352-64. DOI:10.1523/JNEUROSCI.16-07-02352.1996.
- Mitro P, Hijova E, Mudrakova K. Role of the peripheral serotoninergic system in the pathogenesis of vasovagal syncope. Bratislavskelekarskelisty.2006; 107[6-7]:248-252.
- Berger M, Gray JA, Roth BL. The Expanded Biology of Serotonin. Ann Rev Med.2009;60:355-66.
- Nichols DE, Nichols CD. Serotonin Receptors. Chemical Reviews.2008;108:1614-1641. DOI:10.1021/cr078224o
- 10. Gershon MD, Ross LL. Location of sites of 5-hydroxytryptamine storage and metabolism by radioautography. The Journal of physiology.1966;186[2]:477-492. DOI:10.1113/jphysiol.1966.sp008047
- Ekholm R, Ericson LE, Lundquist I. Monoamines in the pancreatic islets of the mouse. Subcellular localization of 5-hydroxytryptamine by electron microscopic autoradiography. Diabetologia.1971;7[5]:339-348. DOI:10.1007/BF01219468
- 12. Kim H, Toyofuku Y, Lynn FC, Chak E, Uchida T, Mizukami H, Fujitani Y, Kawamori R, Miyatsuka T, Kosaka Y, Yang K, Honig G, van der Hart M, Kishimoto N, Wang J, Yagihashi S, TecottLH, Watada H, German MS. Serotonin regulates pancreatic beta cell mass during pregnancy. Nature medicine.2010;16[7]:804-808. DOI:10.1038/nm.2173

- Kim K, Oh CM, Ohara-Imaizumi M, Park S, Namkung J, Yadav VK, Tamarina NA, Roe MW, Philipson LH, Karsenty G, Nagamatsu S, German MS, Kim H. Functional role of serotonin in insulin secretion in a dietinduced insulin-resistant state. Endocrinology. 2015;156[2]:444-452. DOI:10.1210/en.2014-1687
- 14. Ohara-Imaizumi M, Kim H, Yoshida M, Fujiwara T, Aoyagi K, Toyofuku Y, Nakamichi Y, Nishiwaki C, Okamura T, Uchida T, Fujitani Y, Akagawa K, Kakei M, Watada H, German MS, Nagamatsu S. Serotonin regulates glucose-stimulated insulin secretion from pancreatic beta cells during Proceedings pregnancy. of the National Academy of Sciences of the United States of America.2013;110[48]:19420-19425. DOI:10.1073/pnas.1310953110
- 15. Ohta Y, Kosaka Y, Kishimoto N, Wang J, Smith SB, Honig G, Kim H, Gasa RM, Neubauer N, Liou A, TecottLH, DenerisES, German MS. Convergence of the insulin and serotonin programs in the pancreatic beta-cell. Diabetes.2011;60[12]:3208-3216. DOI:10.2337/db10-1192
- 16. Berger M, ScheelDW, Macias H, Miyatsuka T, Kim H, Hoang P, Ku GM, Honig G, Liou A, Tang Y, Regard JB, Sharifnia P, Yu L, Wang J, Coughlin SR, Conklin BR, DenerisES, TecottLH, German MS. Galphai/ocoupled receptor signaling restricts pancreatic beta-cell expansion. Proceedings of the National Academy of Sciences of the United [PNAS]. 2015;112[9]2888-2893. DOI:10.1073/pnas.131937811
- Kasim SF, HamzahFZ, Al-SharafiNMN. Ameliorative effect of rosemary leaves extract on thyroid gland function in hyperthyroid male albino rats. Biochem. Cell. Arch. 2020; Vol. 20, No. 1, pp. 1241-1246. DOI: 10.35124/bca.2020.20.1.1241

2023

- Al-SharafiNMN, Kasim SF, HamzahFZ. Ameliorative role of PTU and rosemary leaves extract in male rats with hyperthyroidism. EurAsian Journal of BioSciences. 2020; 14, 2353-2359.
- 19. HamzahFZ, Al-SharafiNMN, Kasim SF. Effect of aqueous rosemary extract on some sexual hormones in male rats with high thyroxine level. Iraqi Journal of Veterinary Sciences. 2021; Vol. 35[2]: 369-373
- 20. Brizzi G, Carella C, Foglia MC, Frigino M. Thyroid hormone plasmatic levels in rats treated with serotonin in acute and chronic way. Journal of Physiology-Paris. 1997;91[6]:307-310. DOI:10.1016/s0928-4257[97]82411-4
- 21. Ates O, Cayli SR, Yucel N, Altinoz E, Kocak A, Durak MA, Turkoz Y, Yologlu S. Central nervous system protection by resveratrol in streptozotocin-induced diabetic rats. Journal of clinical neuroscience: official journal of the Neurosurgical Society of Australasia.2007;14[3]:256-260. DOI:10.1016/j.jocn.2005.12.010
- 22. Idris MH, Budin SB, Osman M, Mohamed J. protective role of hibiscus sabdariffa calyx extract against streptozotocin induced sperm damage in diabetic rats. EXCLI Journal. 2012;11:659-669.
- 23. Tietz NW. Clinical Guide to Laboratory Tests, 3rd Ed., W.B. Saunders Company, Philadelphia. 1995;509-580. DOI:10.1111/j.1537-2995.1995.tb03571.x
- 24. Temple RC, Clark PM Hales CN. Measurement of insulin secretion in type 2 diabetes: problems and pitfalls. Diabetic Medicine.1992;9[6]:503-512. DOI:10.1111/j.1464-5491.1992.tb01830.x
- Nandi A, Kitamura Y, Kahn CR, Accili D. Mouse models of insulin resistance. Physiological reviews.2004;84[2]:623-647. DOI:10.1152/physrev.00032.2003

- 26. Roep BO, Atkinson M, von Herrath M. Satisfaction [not] guaranteed: reevaluating the use of animal models of type 1 diabetes. Nature reviews. Immunology. 2004;4[12]:989-997. DOI:10.1038/nri1502
- 27. Toye AA, Moir L, Hugill A, Bentley L, Quarterman J, Mijat V, Hough T, Goldsworthy M, Haynes A, Hunter AJ, Browne M, Spurr N, Cox RD. A new mouse model of type 2 diabetes, produced by N-ethyl-nitrosourea mutagenesis, is the result of a missense mutation in the glucokinase gene. Diabetes.2004;53[6]:1577-1583. DOI:10.2337/diabetes.53.6.1577
- Jaim-Etcheverry G, Zieher LM. Electron microscopic cytochemistry of 5- hydroxytryptamine[5-HT] in the beta cells of guinea pig endocrine pancreas. Endocrinology.1968;83[5]:917-923.

DOI:10.1210/endo-83-5-917

29. Lundquist I, Ekholm R, Ericson LE. Monoamines in the pancreatic islets of the mouse. 5-hydroxytryptamine as an modifier intracellular of insulin and the hypoglycaemic secretion. action of monoamine oxidase inhibitors. Diabetologia.1971;7[6]:414-422.

Diabetologia.19/1;/[6]:414-42 DOI:10.1007/BF01212056

- 30. Richmond JE, Codignola A, Cooke IM, Sher E. Calcium- and bariumdependent exocytosis from the rat insulinoma cell line RINm5F assayed using membrane capacitance measurements and serotonin release. PflugersArchiv: European journal of physiology.1996;432[2]:258-269. DOI:10.1007/s004240050132
- Gylfe E. Association between 5hydroxytryptamine release and insulin secretion. The Journal of endocrinology.1978;78[2]:239–248. DOI:10.1677/joe.0.0780239
- 32. Smith PA, Proks P, Ashcroft FM. Quantal analysis of 5hydroxytryptamine release from mouse pancreatic beta-cells. The

Vol. (14) No. (2)

Journal of physiology. 1999;521[Pt3]:651-664. DOI:10.1111/j.1469-7793.1999.00651.x

- 33. Paulmann N, Grohmann M, Voigt JP, Bert B, Vowinckel J, Bader M, Skelin M, Jevsek M, Fink H, Rupnik M, Walther DJ. Intracellular serotonin modulates insulin secretion from pancreatic beta-cells by protein serotonylation. PLoS biology.2009;7[10]:e1000229. DOI:10.1371/journal.pbio.1000229
- 34. DobbersteinLJ, July 24, 2018: Serotonin Linked with Mood, Sleep, Gut Heath, Thyroid, and More, on line article [https://www.wellnessresources.com/n ews/serotonin-linked-with-moodsleep-gut-heath-thyroid-and-more]
- 35. Reed TT. Lipid peroxidation and neurodegenerative disease. Free radical biology and medicine.2011;51[7]:1302-1319. DOI:10.1016/j.freeradbiomed.2011.06 .027
- 36. Sultana R, Perluigi M, Butterfield DA. Lipid peroxidation triggers neurodegeneration: a redox proteomics view into the Alzheimer disease brain. Free radical biology and medicine.2013;62:157-169. DOI:10.1016/j.freeradbiomed.2012.09 .027
- 37. Kim GH, Kim JE, RhieSJ, Yoon S. The role of oxidative stress in neurodegenerative diseases. Experimental neurobiology.2015;24[4]:325-340. DOI:10.5607/en.2015.24.4.325

- Arimon M, Takeda S, Post KL, Svirsky S, Hyman BT, Berezovska O. Oxidative stress and lipid peroxidation are upstream of amyloid pathology. Neurobiology of disease.2015;84:109-119. DOI:10.1016/j.nbd.2015.06.013
- 39. Andorn AC, Pappolla MA. Catecholamines inhibit lipid peroxidation in young, aged, and Alzheimer's disease brain. Free radical biology and medicine. 2001;31[3]:315-320.

DOI:10.1016/s0891-5849[01]00580-9

- 40. Herraiz T, Galisteo J. Endogenous and dietary indoles: a class of antioxidants and radical scavengers in the ABTS assay. Free radical research. 2004;38[3]:323-331. DOI:10.1080/1061186031000164816 7
- 41. Horáková L, Ondrejicková O, Bachratá K, Vajdová M. Preventive effect of several antioxidants after oxidative stress on rat brain homogenates. General physiology and biophysics. 2000;19[2]:195-205.
- 42. Ng TB, Liu F, Zhao L. Antioxidative and free radical scavenging activities of pineal indoles. Journal of neural transmission [Vienna, Austria:1996]. 2000;107[11]:1243-1251. DOI:10.1007/s007020070014
- 43. Cadenas E, Simic MG, Sies H. Antioxidant activity of 5hydroxytryptophan, 5-hydroxyindole, and DOPA against microsomal lipid peroxidation and its dependence on vitamin E. Free radical research communications.1989;6[1]:11-17. DOI:10.3109/10715768909073423