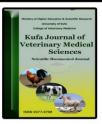
Kufa Journal for Veterinary Medical Sciences Vol.(5). No.(2) 2014



Kufa Journal for Veterinary Medical Sciences www.vet.kufauniv.com



# Effect of ferulic acid on Myocardial Ischemia/Reperfusion Injury In Male Mice.

Bassim I. Mohammad AL-Sheibani<sup>\*</sup> Najah Raiesh Hadi \*\* Najwan Abdulameer Atia <sup>\*\*</sup> Fadhil Ghaly. Yousif <sup>\*\*\*</sup>

<sup>\*</sup> Department of Pharmacology and Therapeutics, College of pharmacy, Qadisiyah University, Iraq

<sup>\*\*</sup> Department of Pharmacology and Therapeutics, College of Medicine, Kufa University, Iraq

\*\* Department of cardiovascular surgery, College of Medicine, Kufa University, Iraq

### Abstract

Myocardial ischemia–reperfusion represents a clinically relevant problem associated with thrombolysis, angioplasty and coronary bypass surgery. Injury of myocardium due to ischemia–reperfusion includes cardiac contractile dysfunction, arrhythmias as well as irreversible myocytes damage.

A total of 24 Swiss albino adult male mice were randomized into 4 equal groups. Group (1) sham group; Group (2) control group; Group (3) control vehicle (DMSO) group; Group (4) Ferulic acid treated group.

Results of this study showed that levels of myocardial IL-6, were increased (p<0.001) in the control group. Histologically, all mice in control group showed significant cardiac injury (p<0.001). Ferulic acid significantly counteract the increase in myocardium level of IL-6 (P < 0.001). Histological analysis revealed that ferulic acid markedly reduced (P < 0.001) the severity of cardiac injury in the mice underwent LAD ligation procedure.

**Keywords**: Ischemia-reperfusion (I/R) ,systemic inflammatory response syndrome (SIRS), multiple organ dysfunction syndrome (MODS) , Ferulic acid (FA), left anterior descending (LAD), Dimethylsulfoxide (DMSO).

يمثل نقص ألارواء الدموي لعضلة القلب واعادته مشكلة ذات صلة سريريه مرتبطة بالاحتشاءات و القسطرة و جراحة تغيير الشرايين التاجيه. تشمل إصابة عضلة القلب بسبب نقص التروية الدمويه وإعادتها - ضعف مقلص القلب، عدم انتظام ضربات القلب وكذلك تلف الخلايا العضلية التي لا رجعة فيه. استخدم في هذه الدراسة اربع وعشرون ذكرا بالغا من الفئران البيض السويسرية. وتم توزيعهم بشكل عشوائي الى اربع مجاميع تشمل مجموعة التظاهر ومجموعة السيطرة (غير معالجه) ومجموعة المذيب و المجموعة المعالجة بحامض الفريولك. واظهرت النتائج ان المستوى الى 6-IL قد ازدادت معنويا (p<0.001) في مجموعة السيطرة. أما النتائج النسيجية بعد عملية ربط الشريان التاجي الأمامي النازل الأيسرفقد أظهرت أن جميع قلوب الفئران قد تضررت معنويا(0.001) من خلال نتائج هذه الدراسه نجد بأن حامض الفريولك اظهر تأثيرا معنويا الفران قد تضررت الالتهاب من خلال منع ارتفاع عامل الالتهاب 6-IL معنويا (g<0.001) مي مجموعة السيطرة. أما النتائج معنويا(http://doi.org/10.000) من خلال نتائج هذه الدراسه نجد بأن حامض الفريولك الإليهاب من خلال منع ارتفاع عامل الالتهاب 6-IL معنويا الأيسرفود أظهرت أن جميع قلوب الفئران قد تضررت الالتهاب من خلال منع ارتفاع عامل الالتهاب 6-IL معنويا الأسريولك الفر تأثيرا معنويا (g<0.001) ملى الالتهاب من خلال منع ارتفاع عامل الالتهاب 6-IL ما بالنسبة النتائج النسيجية بينت ان حامض الفريولك اظهر انخفاضا معنويا (g<0.001) بحدة الضرر الناتج بالفئران نتيجة عملية ربط الشريان التاجي الأمامي النازل الأيسر المسببة للإصابة بالنقص القلبي بالإرواء الدموي/ اعادة الارواء.

### **Introduction:**

Myocardial ischemia is a condition in which blood flow and oxygen to the cardiac muscle is decreased by partial or complete blockage of an artery carrying blood to the myocardium leading to death of an affected cardiac muscle, while the restorations of blood flow to an ischemic heart refer to myocardial reperfusion (1).

Reperfusion injury is the tissue damage caused when blood supply returns to the tissue after a period of ischemia or lack of oxygen (2). The absence of oxygen and nutrients from blood during the ischemic period creates a condition in which the restoration of circulation results in inflammation and oxidative damage through the induction of oxidative stress rather than restoration of normal function (3).

Ischemia and reperfusion injuries (I/R) are a primary cause of cardiac failure, morbidity, and mortality after cardiac operations (4) and heart infarctions (5).

I/R injury represents a fundamental common pathway of tissue damage in a wide variety of disease and surgical processes such as major trauma, acute mesenteric ischemia, septic and hypovolemic shock, abdominal aortic aneurism surgery, and cardiopulmonary bypass (6). Interruption of blood supply results in ischemic injury to all body systems and especially to high metabolically active tissues (7). Paradoxically, restoration of blood flow to the ischemic tissue augments cell injury by delivering mediators induced in toxic the ischemic tissue into the circulation thus affecting distant organs. This might lead to the development of systemic inflammatory response syndrome (SIRS), which can progress multiple to organ dysfunction syndrome (MODS) (6).

I/R injury promotes a release of oxygen free radicals, cytokines and proinflammatory other mediators that activate both the neutrophils vascular the coronary and endothelium. Activation of these cell types promotes the expression of adhesion molecules on both the neutrophils endothelium, and which recruits neutrophils to the surface of the endothelium and initiates a specific cascade of cellcell interactions, leading first to adherence of neutrophils to the vascular endothelium, followed later by trans endothelial migration and direct interaction with myocytes (8).

I–R injury may occur in a variety of clinical settings, including reperfusion after thrombolytic therapy, coronary angioplasty, organ transplantation, aortic cross-clamping or cardiopulmonary bypass (9). Ferulic acid a 4-hydroxy-3-methoxycinnamic acid isolated from the root of *S.buergeriana*, is a kind of phenolic acid that is abundant in many of herbal medicines (10).

FA is a ubiquitous phenol in seeds and leaves of various plants and an antioxidant found in many staple foods, such as fruits, vegetables, cereals, coffee and in plant constituent exhibiting a wide range of therapeutic effects such as anticancer, antidiabetic, cardio protective and neuroprotective, anti-inflammatory activity (11).

**Chen** *et al.*(2009) found that salt of FA can inhibit myocardial cell death after anoxia/reoxygenation by reducing Ca2+ overload. However, the molecular target of FA in cell death signaling during I/R has not been fully identified (12).

Ferulic acid resist platelet can aggregation. inhibit platelet 5hydroxytryptamine release, and inhibit platelet thromboxane A2 (TXA2) generation. It can reduce myocardial ischemia and oxygen consumption (13) and is used in clinical practice to treat coronary heart disease and angina pectoris mainly through inhibiting lipid oxidation, lowering cholesterol content in serum and antithrombotic effects. It can prevent atherosclerosis so as to treat coronary heart disease (14).

### **2-Materials and Methods**

### 2.1. Animals

A twenty-four adult males Swiss Albino mice weighing 28-33 g were purchased from Animal Resource Institute of embryo Center. the research and treatment of infertility, Al-Nahrain University, Iraq. The animals were apparently healthy and they were housed in the animal house of College of Medicine/University of Kufa in a temperature-controlled (25± 2°C) room with alternating 12-h light/12-h dark cycles and were allowed free access to water and chow diet until the start of the experiments. The study was approved by the ethical commitments of institution.

### 2.2.In vivo myocardial I/R model

The *in vivo* myocardial I/R model was modified from a previous study (15). Briefly, mice were anesthetized with with 100mg/kg ketamine and 10mg/kg xylazine (16). The mice were intubated Mechanical ventilation is then and achieved by connecting the endotracheal tube to scientific ventilator (Harvard Model) at a respiratory rate of 138 breath/minute with a tidal volume of 20 mL/kg body weight. A left thoracotomy was carried out to expose the heart. The LAD is then transiently ligated (or can be tied with a slipknot) using a 6-0 polyprolene suture for a 30-minute ischemic period (17). After a 30-min ischemia, by Microsurgical scissors are used to cut the knot in the ligature (or by releasing the slipknot) the heart was reperfused for 2 h. Immediately after finishing the reperfusion time the mouse was sacrificed ,starting by injection of high dose from ketamine and xylazine, after giving good time for the animal to go into deep anesthesia, the mouse is positioned and the chest is opened in flap like manner revealing the heart then a needle of the syringe is introduced into right ventricle to aspirate around 2 ml of blood for later plasma analysis. After that hearts were rapidly removed for quantification of myocardial injury by biochemical studies.

### 2.3.Experimental groups and protocols

After the two week of acclimatization the mice were randomized into 4 groups, 6 mice in each group as follow:

- 1. <u>Sham group</u>: mice underwent the anaesthetic and surgical procedures but without left anterior descending (LAD) coronary artery occlusion.
- <u>Control group</u>: (induced untreated group) mice underwent LAD coronary artery occlusion (for 30 min.), then reperfusion for 2 hours.
- <u>Control vehicle (DMSO) treated</u> <u>group</u>: mice pretreated with Dimethylsulfoxide (DMSO) 1% v/v in normal saline i.p. at 1hr. before ischemia.
- 4. <u>Ferulic acid treated group (FA)</u>: mice pretreated with FA 50

mg/kg i.p. at 1 hr. before ischemia.

### 2.4.Tissue Preparation for measurement of (IL-6)

Cardiac tissues collected 120 minutes after reperfusion were homogenized in solution containing a 1:10(w/v)phosphate buffered saline that contain 1% triton X-100 and protease inhibitor cocktail (18).by using high intensity liquid processor. After homogenization, samples were centrifuged at 14,000 rpm for 20 min at the supernatant was  $4^{\circ}C$  (19) collected used and in IL-6 measurement commercially using available ELISA kits (Bioscience) according to the manufacturer's instructions.

## 2.5.Histopathological Analysis and Damage Scoring

Tissues fixed 4% were in paraformaldehyde and embedded in paraffin. Sections were stained with hematoxylin and eosin for histological evaluation of tissue damage. In order to have a quantitative estimation of cardiac damage, sections (n=6 for each animal) were scored by 2 independent observers blinded to the experimental protocol. The following morphological criteria were considered: score 0, no damage; score 1 (mild), interstitial edema and focal necrosis; score 2 (moderate), diffuse myocardial cell swelling and necrosis; score 3 (severe), with necrosis the presence of contraction bands and neutrophil infiltrate; and score 4 (highly severe), widespread necrosis with the presence contraction bands, neutrophil of infiltrate, and hemorrhage.

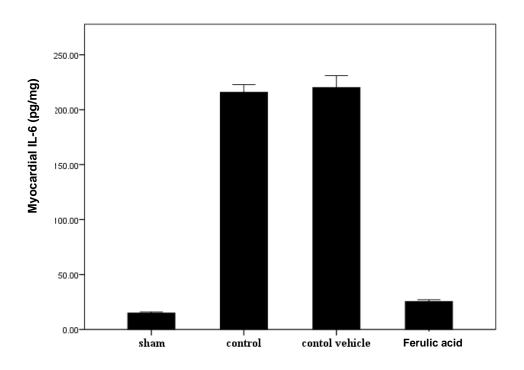
#### 2.6. Statistical Analysis

Statistical analyses were performed using cytokines SPSS 20.0 for windows.lnc. Data were analyzed by ELISA. Figure(1) shows expressed as mean ± SEM. Analysis of that MI/R injury increased significantly Variance (ANOVA) was used for the (p<0.001) the levels of myocardium multiple comparisons among all groups IL-6 compared with the sham group followed by post-hoc tests using LSD (P < 0.001). In the method.

### **3. RESULTS**

3.1. Ferulic acid reduced myocardial IL-6 following MI/R injury

Myocardium levels of inflammatory following MI/R were ferulic acid treatment group, myocardium levels of IL-6, were reduced significantly compared with the control group (P <0.001).



Figure(1): The mean of myocardial IL-6 level (pg/mg) in the four experimental groups at the end of the experiment

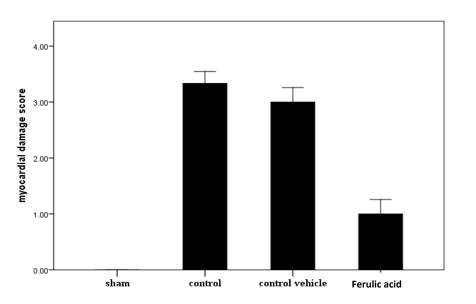
### **3.2.** Histopathological findings

Treatment of mice with Ferulic acid improved cardiac injury significantly (P < 0.001) as compared with control vehicle group and the total severity scores mean of this group showed that 83.3% had mild cardiac injury and 16.7% had moderate cardiac injury. A cross section of sham mouse's heart showed a normal cardiac structure. All mice in this group showed normal hearts 100% as shown in table(1). There was statistically insignificant difference between control group and control vehicle group (P>0.001) and the total severity scores of the control vehicle group showed 16.7% of the group had moderate, 66.7% had severe and 16.7% had highly severe cardiac injury.

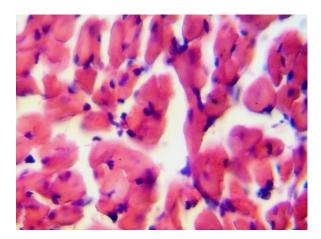
 Table (1): The differences in histopathological scoring of abnormal heart

|                         | Sham group |      | Control group |       | Control Vehicle<br>group |       | Ferulic acid<br>treated group |       |
|-------------------------|------------|------|---------------|-------|--------------------------|-------|-------------------------------|-------|
| Normal                  | 6          | 100% | 0             | 0     | 0                        | 0     | 0                             | 0     |
| Mild                    | 0          | 0    | 0             | 0     | 0                        | 0     | 5                             | 83.3% |
| Moderate                | 0          | 0    | 0             | 0     | 1                        | 16.7% | 1                             | 16.7% |
| Sever                   | 0          | 0    | 4             | 66.7% | 4                        | 66.7% | 0                             | 0     |
| Very Sever              | 0          | 0    | 2             | 33.3% | 1                        | 16.7% | 0                             | 0     |
| Total cardiac<br>injury | 6          | 100% | 6             | 100%  | 6                        | 100%  | 6                             | 100%  |

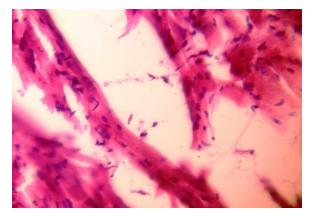
changes among the four experimental groups



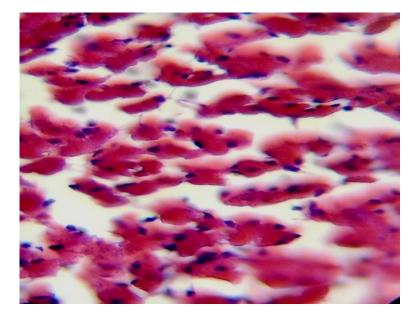
**Figure (2):** Error bar chart shows the difference in mean± SEM values of total severity scores in the four experimental groups.



**Figure (3)**:Photomicrograph of heart section of normal mice shows the normal architecture. The section stained with Haematoxylin and Eosin (X 10).



**Figure (4)**: Photomicrograph of cardiac section for the control vehicle group showed interstitial edema, haemorrhage and PMN infilteration. The section stained with Haematoxylin and Eosin (X 40).



**Figure (5)**: Photomicrograph of cardiac section in ferulic acid treated group. The section show almost normal cardiac tissue, the section stained with Haematoxylin and Eosin (X 40)

#### 4- **Discussion**

The major findings of the present study are as follows. Firstly, that the inflammatory cytokine (IL-6) play important role in the pathology of myocardial I/R Secondly, ferulic acid pretreatment played a protective role against myocardial I/R injury. The protective effects of ferulic acid during myocardial I/R injury were correlated with the attenuation of inflammation and Myocardial damage.

Cardiovascular disease(CVD) with all its appearances and complications is responsible for approximately 50% of all deaths in the world today (20). One of the major contributing factors to CVD and heart failure is the loss of function or indeed death of cardiomyocytes due to coronary vessel occlusion (ischemia) followed by restricted blood flow (reperfusion) to the heart and a corresponding oxygen deprivation due to hypoxia (21). The early reperfusion phase is characterised by enhanced release of ROS from endothelial cells and cardiomyocytes, as well as enhanced expression of cytokines and adhesion molecules, The enhanced expression of chemokines during the first hours of reperfusion triggers further recruitment of neutrophils and monocytes into the

2014

infracted myocardium which lead to increase the cardiac damage by further releasing ROS, inflammatory mediators and proteases (22).

Kim and Lee(2012) indicated that ferulic acid attenuates ischemia/reperfusion-induced hepatocyte apoptosis via inhibition the increases in the serum tumor necrosis factor- $\alpha$  levels and TNF receptor type-1-associated DEATH domain protein and TNF receptor-associated factor-2 protein expressions(23).

Obayashi et al.(2005), Kosmadaki et al.(2003) observed that IL-6 and TNFepidermic α are secreted by keratinocytes and play an important role in UV-mediated photobiology. IL-6 could cause fever and promote the synthesis of C- reactive protein (CRP). There have been reports indicate that immediate adding TNF-  $\alpha$  antibody after UV irradiation reduced cell debris formation and apoptosis remarkably(24,25).

Lin et al.(2010) confirmed that the levels of IL-6 and TNF- $\alpha$  increased 30~90 mJ/cm2 of after UVB irradiation, which further implied the significance of cytokines in UVinduced skin injury. After Ferulic acid treatment, IL-6 and TNF- $\alpha$  secretion was apparently reduced (P <0.001)(26).

### 5-Referances

1- Ansorge S, Reinhold D, Lendeckel U. (2003):Propolis and some of its constituents down-regulate DNA synthesis and inflammatory cytokine production but induce TGF- $\beta$ 1 production of human immune cells. Z Naturforsch. 58c: 580- 89.

No. (2)

2- Grace P. (2005). Ischemia reperfusion injury. British Journal of Surger. 81: 637-47.

3- Manipuri P, Indala R, Jagaralmudi Ramesh Kumar (2013). A. K. Hepatoprotective Effect of Hepatic Inularacemosa on Ischemia/reperfusion Induced Injury in Rats. J Bioanal Biomed 5: 022-027.

4- Basgut B, Kayki G, Bartosova L, Ozakca I, Seymen A, Kandilci H, *et al.*(2010). Cardioprotective effects of 44Bu, a newly synthesized compound, in rat heart subjected to ischemia/reperfusion injury. Eur J Pharmacol 640:117–123.

5- Heusch G, Musiolik J, Gedik N, Skyschally A. (2011) .Mitochondrial STAT3 activation and cardioprotection by ischemic postconditioning in pigs with regional myocardial ischemia/reperfusion. Circ Res 109:1302–8.

6-Leng Y, Zhang Y, Zhang Y, Xue X, Wang T, Kang Y. (2011). Ischemic post-conditioning attenuates the intestinal injury induced by limb ischemia/reperfusion in rats. *Braz J Med Biol Res*, 44(5):411-17.

7- Mallick I, Yang W, Winslet M, Seifalian A. (2004). Ischemiareperfusion injury of the intestine and protective strategies against injury. *Dig Dis ci*, 49(9):1359-77.

8-Jordan J, Zhao Z, Vinten-Johansen J. (1999). The role of neutrophils in

myocardial ischemia-reperfusion injury. Cardiovasc Res. Sep;43(4):860-78.

9-Eltzschig HK, Collard CD .Vascular ischaemia and reperfusion injury. Br Med Bull. 2004 Oct 19;70:71-86.

10- <u>Graf E</u>. (1992). Antioxidant potential of ferulic acid. <u>Free Radic</u> <u>Biol Med.</u> Oct;13(4):435-48.

11- Gohil K, Kshirsagar S and Sahane
R. (2012). Ferulic Acid – A
Comprehensive Pharmacology Of An
Important Bioflavonoidvol. 3(1): 700 - 10.

12- Chen H, Liao Z, Huang Q, He M. (2009). Sodium ferulate attenuates anoxia/reoxygenation-induced calcium overload in neonatal rat cardiomyocytes by NO/cGMP/PKG pathway. Eur. J. Pharmacol. 603:86– 92.

13- Zhang M, Shen Y. (1994). Antihypoxia effect of caffeic acid and ferulic acid. Northwest PharmaceuticalJournal;9:118.

14- Wang H, Shi E. (2007). The advance in the research on phamacology action of ferulic and its derivatives. Acta Academiae Medicinae Neimongol;29:601-3.

15- Kolk M, Meyberg D, Deuse T, Tang-Quan K, Robbins R, Reichenspurner H, *et al.*(2009). LADligation: a murine model of myocardial infarction. <u>J Vis Exp.</u> Oct 14;(32). pii: 1438. 16- Feitoza C, Goncalves G, Semedo P, Cenedeze M, Pinheiro H, Beraldo F, *et al.*(2008). Inhibition of COX 1 and 2 prior to renal ischemia/reperfusion injury decreases the development of fibrosis. Mol Med 14(11-12): 724-30. 17- Maekawa N, Wada H, Kanda T, Niwa T, Yamada Y, Saito K, *et al.*(2002). Improved myocardial ischemia/reperfusion injury in mice lacking tumor necrosis factor-alpha. J Am Coll Cardiol. Apr 3;39(7):1229-35.

2014

18-Zhang M, Xu YJ, Saini HK, Turan B, Liu PP, Dhalla NS.Pentoxifylline attenuates cardiac dysfunction and reduces TNF-alpha level in ischemic-reperfused heart.Am J Physiol Heart Circ Physiol.(2005 b)Aug;289(2):H832-9. Epub 2005 Apr 15.

19-Granado M, Fernández N, Monge L, Figueras JC, Carreño-Tarragona G, Amor S, García-Villalón AL. Effects of coronary ischemia-reperfusion in a rat model of early overnutrition. Role of angiotensin receptors. PLoS One. 2013;8(2):e54984. doi: 10.1371/journal.pone.0054984. Epub 2013 Feb 1.

20-Allender S, Scaraborough P, Peto V. (2008). European cardiovascular disease statistics. 2008 edition.

21- Barry S, Townsend P, Latchman D, Stephanou A. (2007). Role of the JAK-STAT pathway in myocardial injury. Trends Mol Med; 13(2):82-89.

22-Steffens S, Montecucco F, Mach F .The inflammatory response as a target to reduce myocardial ischaemia and

2014

reperfusion injury. Thromb Haemost. 2009 Aug;102(2):240-7.

23-<u>Kim H, Lee S</u>. (2012). Ferulic acid attenuates ischemia/reperfusioninduced hepatocyte apoptosis via inhibition of JNK activation. <u>Eur J</u> <u>Pharm Sci.</u> Apr 11;45(5):708-15.

24- Obayashi K, Kurihara K, Okano Y, Masaki H, Yarosh D. (2005):L-Ergothioneine scavenges superoxide and singlet oxygen and suppresses TNF-alpha and MMP-1 expression in UV-irradiated human dermal fibroblasts. J Cosmet. Sci. 56(1): 17-27.

25- Kosmadaki MG, Yaar M, Arble BL, Gilchrest BA. (2003): UV induces VEGF through a TNF-alpha independent pathway.FASEB J. 17(3): 446-8.

26- Lin XF , Min W and Luo D. (2010): Anticarcinogenic effect of ferulic acid on ultraviolet-Birradiated human keratinocyte HaCaT cells. Journal of Medicinal Plants Research. 18 August Vol. 4(16): pp. 1686-1694, ISSN 1996-0875.