Kufa Journal for Veterinary Medical Sciences Vol. (7), No. (2) 2016



Kufa Journal for Veterinary Medical Sciences

vetmed@uoKufa.edu. iq



The Impact of *Eruca sativa* Seeds on Leydig's Cells Number and Hormonal Profile In Cadmium Exposed Rats

Baraa Najim Al-Okaily* and Zenah Mohammed Al-Shammari **

*Department of Physiology and Pharmacology, College of Veterinary Medicine ,University of Baghdad, Iraq. **Department of Biology., College of Basic Education, University of Wassit , Iraq.

Corresponding author : Baraa Najin Al-Okaily

E.mail:Baraanajim@yahoo.com

Abstract

This study was aimed to investigate the protective role of Eruca sativa seeds extract on the number of Leydig's cells and hormonal profile in cadmium chloride treated rats. Forty adult male rats were randomly divided to four equal group and handled as follows for 56 days: Control group administered tap water; group G1 administered tap water containing 30 ppm/ L of cadmium chloride; group G2 received tap water containing 30 ppm/L of cadmium chloride and administered 250 mg/Kg.B.W of ethanolic extract of Eruca sativa seeds; group G3 administered 250 mg/Kg.B.W of ethanolic extract of Eruca sativa seeds only. Collecting of blood samples were obtained at 0, 28 and 56 days for estimation of serum testesterone (T), follicule stimulating hormon (FSH) and leutinizing hormon (LH) concentrations. At the end of experiment body weight and testicular weight were measurment. Furthermore, histological sections of testes were prepared for counting of Leydigs cells. A significant decrease in serum testesterone (T), follicule stimulating hormon(FSH) and leutinizing hormon (LH) concentrations and decrease in body weight to testes weight% in group G1 at concentration of CdCl₂ 30 ppm/ L, as well as, statistical analysis and examination of histological sections of testes revealed a significant decrease in the number of Leydig's cells. Whereas, The results pointed the beneficial effects of Eruca sativa seeds extract to improvement the previous parameters, against cadmium chloride (G2), through a significant increase in hormonal profile concentration and numbers of Leydigs cells parallel with alleviating the testicular toxicity induced by CdCl₂.

Key Words: Cadmium, *Eruca sativa*, Leydigs cells, hormonal profile, rats.

تأثير بذور الجرجير على أعداد خلايا ليدك وصورة الهرمونات في ذكور الجرذان المعرضة للكادميوم

براء نجم العقيلي * و زينه محمد الشمري * * *فرع الفسلجة و الأدويه/كلية الطب البيطري - جامعة بغداد **قسم العلوم/كلية التربيه ألأساسيه - جامعة واسط

الخسلاصيه

أستهدفت هذه الدراسه للتعرف على الدور الوقائي للمستخلص الكحولي لبذور نبات الجرجير في الحد من تأثير كلوريد الكادميوم على عدد خلايا ليدك وصورة الهرمونات في ذكور الجرذان البالغع تم استخدام 40 جرذ بالغ، قسمت إلى أربع مجاميع متساوية وعوملت لمدة 8 أسابيع كالتالي : جرّعت المجموعة الأولى ماء

No. (2)

الشرب واعتبرت مجموعة سيطرة، أما المجموعه الثانية (G1) فقد أعطيت كلوريد الكادميوم بتركيز 30جزء بالمليون التر من ماء الشرب في حين اعطيت المجموعه الثالثة (G2) كلوريد الكادميوم في ماء الشرب كما في الجموعة الأولى أضافة الى تجريعها المستخلص الكحولي لبذور الجرجير بجرعه 250ملغم كغم من وزن الجسم ،بينما جرعت المجموعه الرابعه (G3) بالمستخلص الكحولي لبذور الجرجير بجرعه 250ملغم /كغم من وزن الجسم جمعت عينات الدم للفترات وو 28و 56 يوم من التجربه وتم عزل السيرم لغرض تقدير تركيز هرمون التستستيرون ، الهرمون المحفز للجريب والهرمون الليوتيني في نهاية التجربه وزنت الحيوانات ، وتم أخذ وزن الخصيتين ثم أخذت عينات من الخصية اليمني وعمل لها مقاطع نسيجيه لغرض حساباً عداد خلايا ليدك أظهرت النتائج وجود انخفاض معنوي في معابير الدراسه في الجرذان المعاملة بكلوريد الكادميوم وبتركيز 30جزء بالمليون مقارنة بالمجاميع الأخرى. فضلا عن ذلك أظهرت نتائج الفحص النسيجي حدوث اخفاض معنوي في عدد خلايا ليدك للجر ذان المعاملة بكلوريد الكادميوم. وعلاوة على ذلك، فقدأشارت النتائج الى الدور الأيجابي "المستخلص الكحولي لبذور الجرجير في تقليل حدة الأثار الضارة لكلوريد الكادميوم وتحسين المعايير المدروسه من خلال الزيادة المعنوية في تركيز الهرمونات وزيادة اعداد خَلَايًا ليدك يُنستَنتج من ذلك أن المستخلص الكحولي لبذور الجرجير يمتلك خصائص مضادة لتقليل تسسم الخصى عن كلوريد الكادميوم. الكلمات المفتاحيه: كادميوم، الجرجير، خلايا ليدك، صورة الهرمونات، الجرذان

Introduction

The main natural sources cadmium include volcanic activity, sea spray, soils, sediments and from rain getting into our water supplies (1), besides, using of phosphate fertilizers will increasing the compounds of cadmium in ecosystem(2). in Occupational exposure metal industries (3) and nonsmoking population consider as a sources for Cd toxicity (4). At early date cadmium was proved to be induced physiological disruption of spermatogenesis and damage sperm DNA (5) leading to decline of fertility in mammalian species. Cadmium affect male reproductive the system particularly testes causing testicular degeneration (6). However, several studies suggested that exposure to Cd caused spermatological damage and histological alteration in testes epididymis and accessory glands with disruption blood-testes-barrier of (BTB) leading to testicular toxicity and reduced testicular size (6,8 and 9).

Some research stated that cadmium can induce neuroendocrine toxicity by disrupting the pattern of hypothalamicpituitary –gonadal axis in male leading to acquired malformations such as cryptorchidism, as well as, testicular

cancer may occur (10). has ability to generate organisms reactive oxygen species (ROS) in responses to cadmium exposure (11). Over-accumulation of ROS leads to oxidative stress (OS) which, in turn, causes oxidative damage of proteins and DNA leading to increased rate of mutations as well as, lesions in various biological molecules such peroxidation of lipids (LPO) (12). Besides ROS caused an apoptosis in the testis may be mediated through alterations of mitochondrial apoptotic pathway (13).Nevertheless. spermatozoa can generate normally within limited levels for its normal functions(14). Whereas, excess production of ROS in some conditions such as in cadmium toxicity caused adversely effect on sperm functions (15). Consequently, the presence of dysfunctional spermatozoa in the semen caused a significantly elevation of ROS production (16). It has been known that antioxidants protect the cells directly or indirectly from the damage caused by toxic radical reactions through different mechanisms 18).*Eruca* sativa and commonly known as rocket. The phytochemical composition of Eruca seeds and revealed the existence of glucosinolates such allyl as

sulphocyanate (19),in addition, isolated and identified three new types of quercetins derivatives from its E.sativa leaves(20). Aliva, (21) found that (4 methylthiobutylisothiocyanate) Erucin and are the isothiocyanates in E. sativa seeds . Evidently in alternative medicine, rocket had been used as medicinal plant for different purpose due its antioxidant properties (22,23 24). Considering these phytochemical compounds have an antioxidant activities, so most attention has been focused on its seeds, therefore, this experiment was designed to investigate the effect of alcoholic extract of Eruca sativa seeds on hormonal profile and Leydigs cells against cadmium chloride in adult rats.

Materials and Methods

Eruca sativa seeds were purchased from the local market (Shorje -Baghdad). The ethanolic extract of Eruca sativa was prepared as described by(25) as follows: 100 g of Eruca sativa seeds were ground into powder then put it in a volumetric conical flask 1000 ml of 70 % ethyl alcohol was added to the powder and mixed by using magnetic stirrer apparatus for 24hr, filtration of the mixture by using 4 layers of medical gauze, then was filtered again using Whatman (No.1) filter paper. The filtered mixture was evaporated by using incubator at 40°C for 72hr to obtain crude extract. The yield equal to 3g.

Forty adult mature - week old male rats were randomly divided into four equal groups and one group was considered as control group ,whereas, the other groups were handled as follows for 56 days : group G1 administered tap water containing 30 ppm/ L of cadmium chloride; group G2 received tap water containing 30 ppm/ L of cadmium chloride and

administered 250 mg/Kg.B.W ethanolic extract of Eruca sativa seeds administered G3 group 250 mg/Kg.B.W of ethanolic extract of Eruca sativa seeds only. Collecting of blood samples were obtained via cardiac puncture from each anesthetized animal at 0, 28 and 56 days of the experiment then centrifuged at 3000 rpm for 15 minutes, and sera was isolated and frozen at -18 Co till analysis of serum testesterone (T), follicule stimulating hormon(FSH) and leutinizing hormon concentrations (LH) by using immunoenzymometric assay Kits (Monobind Inc, USA). Body weight and testicular weight were estimated. Furthermore, testis were excised and fixed in 10% formalin buffer solution for histological examination. were embedded in paraffin blocks and several tissue sections with thickness of 6 µ was prepared and stained by hematoxylin and eosin according to (26) to study the Leydig cells .Counting of Leydigs cells was done by reading the cells between each three seminiferous tubules,10 cross-section per rat were recorded by using light microscope 40X) (under calculation the mean number Leydigs cells cell / µmm² according to Java-based image processing program (27). Statistical analysis of data was performed on the basis of One-way and Two-Way Analysis of Variance (ANOVA) using a significant level at (P<0.05)and using Least Significant specific Differences (LSD) test for group differences (28).

Results

After 28 and 56 days of the experiment, cadmium treated group (G1) recorded a significant (P<0.05) decrease in serum testosterone concentration as compared to other groups (figure 1-A) . Whereas, the

No. (2)

testosterone concentration in G2 and G₃ groups significantly (P < 0.05)elevated after 28 and 56 days of experiment period comparing to the control and G1 groups. On the other hand, serum testosterone concentration of group G_3 at 28 and 56 days of treatment showed a significant (P<0.05) increase comparing to other groups Besides, treated dependent , a significant (P<0.05) highest level of serum testosterone concentration were recorded in groups G2 and G3 at two experimental periods (28 and 56 days) as compared to zero time.

The data show a marked significant (P < 0.05) reduction in serum FSH concentration in G1 treated group after and 56 days of experiment compared with the others experimental groups (figure 1-B). While, at the end of experiment the results recorded a significant (P < 0.05) increase serum FSH concentration in group G2 (received CdCl₂ plus E.S.) as compared with control and G1 groups. Besides, there was no significant (P> 0.05) differences in FSH concentration between G2 and G3 at the end of the experiment when compared among them.

Figure 1-C illustrates the mean value of LH hormone. In comparison between groups the results showed a general trend for the LH value to decrease significantly (P<0.05) 56 days in Cd treated group (G1) as compared to control and other treated groups. In addition, the results showed that rats received CdCl₂ concurrently with Eruca sativa extract (group G2) exhibited a significant (P < 0.05) increase in LH at days 28 and 56 days of the experiment comparing to group G1 . It was also noted the existence of a significant (P<0.05) elevation in serum LH concentration in group G3 (received E.S.) as compared to control and G1 groups. Within the time, with exception to group G3, the results recorded absence of statistical (P>0.05)differences in all groups the experimental period along comparing to the pretreatment period.

Data pertaining, in figure 2-A, recorded a significantly (p<0.05) decrease in testicular weight to body weight ratio in Cd treated group (G1) as compared with the control, G2 and G3 groups. On the contrary, the results showed a significant(p<0.05) increase this parameter group G3 as compared with the control .G1 and G2 groups. It was also noted the absence of statistical differences between the two G2 and G3 when compared with each other. Statistical analysis of the number of Leydig's cells showed a significant decrease (P<0.05) in rats exposed to CdCl2 at concentration 30Pppm/L at end of the experiment (figures 2-B, 4-A and 4-B) compared to other groups. Besides, the histological section of testes of rats received CdCl₂ plus E.S extract (group G2) revealed a significant increase (P<0.05) the number of Leydig's cells (figures 2-B and 5) as compared to G1 (figures 4-A and 4-B). On the other hand, the number of Leydig's cells of ES treated group only (figures 2-B and 6) showed significant (P<0.05) increase in this parameter as compared with control, G1 and G2 groups (figures 3-B, 4-B and 5).

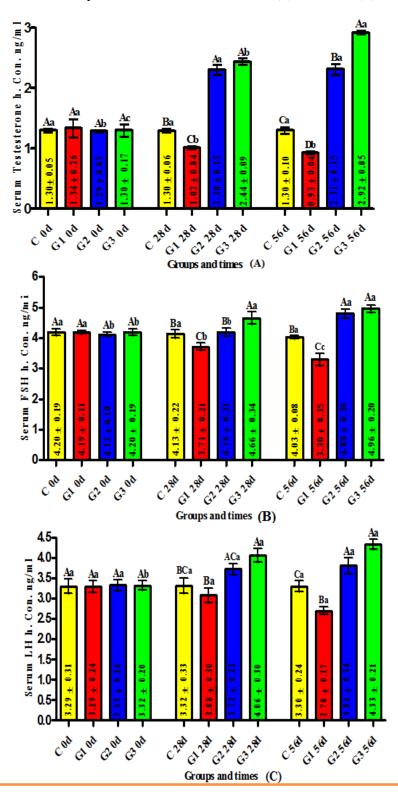


Figure.(1): Effect of alcoholic extract of Eruca sativa seeds and cadmium chloride on hormonal testosterone (A), FSH (B) and LH (C) concentrations in adult male rats exposed to cadmium chloride.

Value are expressed as mean \pm SE, n=10 each group

C: control, G1: administration of 30 ppm/L of CdCl2. G2: administration 250 mg/kg B.W. ethanolic extract of Eruca sativa seeds plus 30 ppm/L of CdCl2 . G3: administration 250 mg /kg B.W. ethanolic extract of Eruca sativa seeds.

Capital letters denote differences between groups P<0.05 Vs. control.

Small letters denote differences within group P<0.05 Vs. control.

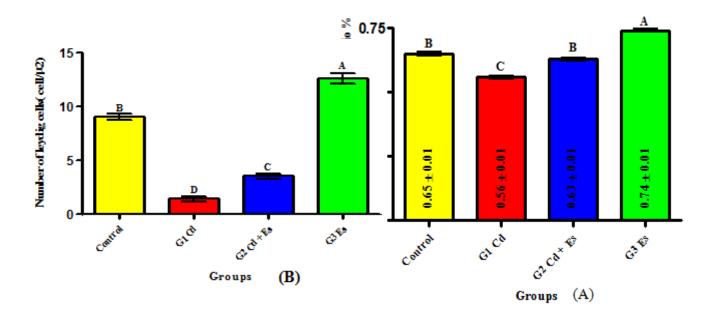


Figure .(2): Effect of Eruca sativa extract and cadmium chloride on testicular weight to body weight ratio (A) and on number of Leydig cells (B) in adult male rats.

Value are expressed as mean \pm SE, n=10 each group

C: control, G1: administration of 30 ppm/L of CdCl2. G2: administration 250 mg/kg B.W. ethanolic extract of Eruca sativa seeds plus 30 ppm/L of CdCl2 . G3: administration 250 mg/kg B.W. ethanolic extract of Eruca sativa seeds. Capital letters denote differences between groups P<0.05 Vs. control.

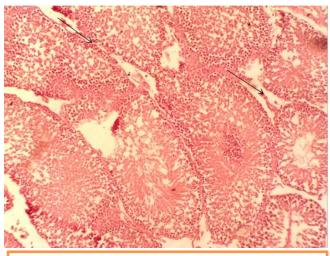


Figure. (3- A): Cross section of rat testis of control group . note :- leydig cells with normal structure \longrightarrow (H &E stain 10 X)

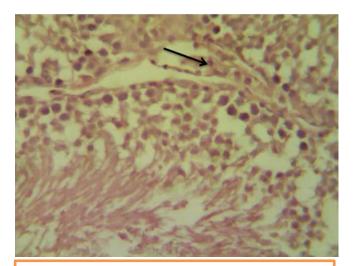


Figure.(3-B): Cross section of rat testis of control group . note:- leydig cell \rightarrow with normal structure (H &E stain 40 X)

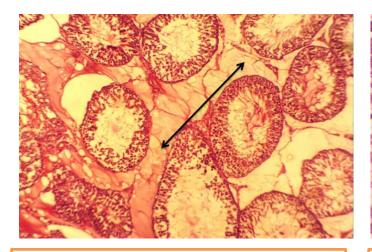


Figure.(4-A):Cross section of rat testis of group G1 treated with CdCl2 (30 ppm/L) for 56 days . noteshow vacculation \iff and absent of leydig cell (H &E stain 10 X).

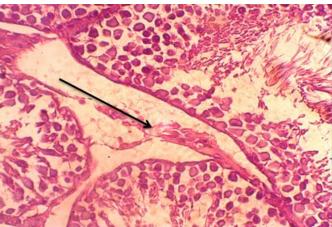


Figure. (4-B):Cross section of rat testis of group G1 treated with CdCl2 (30 ppm/L) for 56 days . note :- few numbers of leydig cells \longrightarrow (H &E stain 40 X)

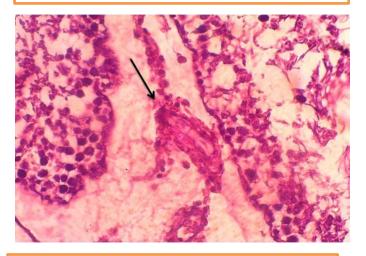


Figure.(5): Cross section of rat testis of group G2 treated with CdCl2 plus 250 mg of *Eruca Sativa* extract for 56 days . note :- show proliferation of leydig cell —>(H &E stain 40 X).

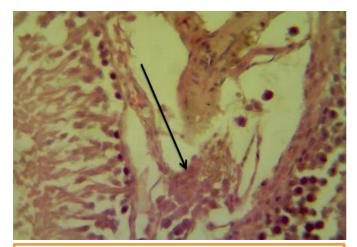


Figure.(6): Cross section of rat testis of group G3 treated with 250 mg of Eruca Sativa extract for 56 days . note :- show normal proliferation of leydig cell \longrightarrow (H &E stain 40 X)

Discussion

Testosterone is a vital male sex hormone produced by Leydig cells for maintenance spermatogenesis process along with FSH (29). Indeed, the result of this study finding that treatment of rats with cadmium chloride at concentration of 30 ppm/L (G1) caused decrement in hormonal profile including testosterone, FSH and LH concentrations and numbers of Leydig cells in comparison with other groups. Many studies manifested that exposure of rats to Cd induced oxidative stress through stimulating the synthesis of cadmium binding proteins (metallothioneins) and heat proteins (30). This decrement may be due to oxidative damage induced by cadmium chloride (11). Asadi et al., (31) reported that Cd caused inhibition of the serum levels of FSH and LH which induced the signals inhibition of testosterone synthesis led to reproductive dysfunction, cell death and apoptosis. The reduction of testosterone level in G1 treated rats may be due to reduction or impaired the utilization of cholesterol by the Leydig cells (32). Moreover, any chemical agent impaired or suppressed the secretion of pituitary gonadotropins will produce antifertility effects with deficient androgen secretion by the testes, cessation of spermatogenesis and loss of libido (33 and 34).

Cd has been considered as an important environmental endocrine disrupter (35), this may leads to variations or modifying the hormones secreted daily by the pituitary gland (36). Elgawish and Ghanem (37), also noted that Cd cause devastation the hypothalamus-pituitary gonadal axis thus, destroying the secretory organs of hormones and compromising Thereby, hormonal gland disruption of this axis may contribute to a reduction in gonadotropins level FSH and LH leading to inhibition of steroid biosynthesis (38). It is well known that small amounts of reactive oxygen species (ROS) are produce normally in the steroidogenic pathway are necessary for fertilizing capabilities (14). Also (13) reported that exposure to Cd could induced oxidative stress through decrease the activities of SOD, GPx and CAT with increasing Cd concentrations, which was accompanied with the increase in malondialdehyde (MDA) and H₂O₂ and hydroxyl radicals content in a concentration-dependent manner and these radicals promote oxidative damage as well as apoptosis in the testis, and the apoptotic processes may mediated via mitochondriadependent apoptosis pathway regulating the activities of caspase-3 and caspase-9 (17).

From the results, it seem that alleviating of hormonal profile in G2 and G3 treated groups by ethanolic extract of E. sativa may be due to its antioxidant defense mechanism and free radicals scavenger activity (39) of phytochemical compounds including glucosinolate and flavonoids which possess a potent antioxidant activity (19 and 40) against oxidative damage induced by Cd and thereby improving the pituitary-testes axis and fertility(41 and 42). E. sativa seeds extract exhibited an evidence for a stimulatory effects on reproductive gonadal system through androgenic activities through increase the number of Leydig cells in G2 and G3 groups could be due to the free radical scavenging ability through excluding the Fe3+ (43 and 44), or may be due to an increasing the number and/or the sensitivity of receptors of the Leydig cells to LH which led to

No. (2)

increase testosterone biosynthesis (45). According to the results exposure of rats to CdCl2 at concentration 30 ppm/L caused decreased in body weight to testicular weigh ratio as compared to other groups, which are in agreement with other studies (32 and 46). These changes suggested that ROS induced and released from cadmium such as hydrogen peroxide from anions and peroxide of hydrogen radicals leading to OS and testicular degeneration (47). Adamkovicova and his collages(48) stated that Cd induced testicular pathogenicity includes edema, cellular degeneration, necrosis of testicular tissue and leading to infertility (6) . Our findings in this study showed a decrease in testicular weight to body weight ratio in Cd treated rats may be due to a possible alteration in androgen status most likely due to a decrease in testosterone, these results are in line with (49). Also a significant decline of gonadotropins depicted in the present study might indicate due to testicular necrosis and degeneration seminiferous tubules as a result of contraction of myoid cells (50) or a change in the proportion of collagen mediated Cd toxicity can affect the structures of testis(46,51 and 53) and contributed to weight loss of testis.

On the other hand, Eruca extract was effective in ameliorating most of the toxic effects induced by Cd on body and testicular weights in G2 and G3 groups as compared to G1. It is well known that maintenance of the testicular weight with the accessory reproductive glands depends testosterone level (53). Accordingly, the significant increase in weights of testis of rats that treated with ES may be due to an improvement in testosterone and / or FSH levels or may be due to the

anabolic effects of testosterone leading to raising the body weight (54), also this findings are in agreement with (41) who revealed that administration of Eruca sativa extract protected the testis of rats exposed to hydrogen peroxide evidenced by appearance of normal structures of seminiferous tubules of testis and preserve spermatogenesis in addition, testosterone could increase the number and functions of somatic and germinal cells of testis followed by testes weight improvement (55). This increment may be due to the effects of ES phytochemical substances in ES including; proteins, vitamins like A, and C, in addition to presence of important minerals (21 and 56). Therefore, treatment of rats with ES extract can protect pituitary-testicular axis and preserve hormonal profile from the detrimental effects of Cd but its effectiveness is dependent on the dose of Cd exposed.

References

- 1-Duruibe, J.O.; Ogwuegbu, M.O.C. and Egwurugwu, J.N. (2007). Heavy metal pollution and human biotoxic effects. Int. J. Phys. Sci., 2(5): 112-118.
- 2-Ayers, R.U.(1992). Toxic heavy metals: materials cycle optimization. Pro Natl Acad Sci., USA. 89: 815-820.
- 3-ATSDR (Agency for Toxic Substances Registry), Disease (1999).and Toxicological Profile for Cadmium (Final Report). NTIS Accession No. PB99-166621. P: 434.
- 4-Pizent, A.; Tariba, B.; and Zivkovic, T.(2012). Reproductive toxicity of metals in men. Arh Hig Rada Toksikol., 1: 35-46
- 5-Bench, G.; Corzett, M.; Martinelli, R.; and Balhorn, R.(1999). Cadmium concentrations in the testes, sperm, and spermatids of mice subjected to long term cadmium chloride exposure. Cytometry.,35(1): 30-36.

- 6-Mohamed, D.; Saber, A.; Omar, A. and Soliman, A. (2014) . Effect of Cadmium on the Testes of Adult Albino Rats And The Ameliorating Effect of Zinc and Vitamin E. British Journal of Science., 11 (1): 72-95.
- , M.M.; Khaled , M.A.; Hassanein , K.M..A. and Waleed , S.W. (2014). Protective effects of thymoguinone and I-cysteine cadmium - induced toxicity in rats . Toxicol Reports ., (1): 612 - 620.
- 8-Marettová, E.; Maretta, M. and Legáth, J. (2015). Toxic effects of cadmium on testis of birds and mammals: a review. Anim Reprod Sci. ., 155: 1-10.
- 9-Minutoli, L.; Micali, A.; Pisani, A.; Puzzolo.D.: Bitto.A.: Rinaldi.M.: Pizzino,G.; Irrera, N.; Galfo, F.; Pallio, G.: Mecchio, A.: Arena.S.: Germana, A.; Bruschetta, D.; Laur, R.; Magno, L.; Marini, H.; Squadrito, F. and Domenica Altavilla, D.(2016).Flavocoxid Protects Against Cadmium-Induced Disruption of the Blood-Testis Barrier and Improves Testicular Damage and Germ Cell Impairment in Mice. Toxicol Sci., 149(1):270.
- 10- Lafuente, A. (2013). The hypothalamic-pituitary-gonadal axis is target of cadmium toxicity. An update of recent studies and potential therapeutic approaches. Food and Chemical Toxicology 59: 395-404.
- 11- Vestena, S.; Cambraia, J.; Ribeiro, C.; Oliveira, J.A. and Oliva. (2011).Cadmium induced oxidative antioxidative stress and enzyme response in Water Hyacinth and Salvinia. Braz J Plant Physiol., 23: 131–139.
- 12- Valko, M.; Rhodes, C.J.; Moncol, J.; Izakovic, M. and Mazur, M.(2006). Free radicals, metals and antioxidant in oxidative stress-induced cancer. Chem Biol Intercat., 106: 1'-40.

13- Wang, L.; Xu, T.; Lei, W.; Liu, D.; Li, Y.; Xuan, R. and Ma, J.(2011). Cadmium-induced oxidative stress and apoptotic changes in testis of fresh water Crab, Sinopotamon henanense. PloS One . 6(11): e27853. Doi: 10.1371/journal.pone.0027853.

No. (2)

- 14- Agarwal, A.; Virk, G.; Ong, C. and du Plessis, S.S.(2014). Effect of Oxidative Stress Male on Reproduction .World J Mens Health .,32(1): 1-17.
- 15- Hampl, R.; Drabkova, P.; Kandar, R. and Stepan, J.(2012).Impact of oxidative stress on male infertility. Ceska Gynekol.,77:241-5.
- 16- Aitken, R.J.; Whiting, S.; De Iuliis, G.N.; McClymont, S.; Mitchell, L.A. and Baker, M.A. (2012). Electrophilic generated aldehydes by mitochondrial metabolism activate reactive oxygen species generation and apoptosis by targeting succinate dehydrogenase. J Biol Chem., 287:33048-60.
- 17- Kara, H.; Cevik, A.; Konar ,V.; Dayangac, A. and Yilmaz, M. (2007). Protective effects of antioxidants against cadmium-induced oxidative damage in rat testes. Biol Trace Elem Res.,120(1-3):205–11.
- 18- Siddique, N.A.; Mujeeb, M.; Naimi, A.K. and Akram, M. (2010). Evaluation of antioxidant activity, quantitative estimation of phenols and flavonoids in different parts of Aegle marmelos. Afr J Plant Sci., 4:1-5.
- 19- D'Antuono, L.F.; Elementi, S. and Neri, R. (2008). Glucosinolates in diplotaxis and Eruca leaves: diversity, taxonomic relations and applied Phytochemistry., aspects. 69:187-199.
- 20- Weckerle, B.; Michel, K.; Balazs, B.; Schreier, P. and Toth, G. (2001). Quercetin 3, 3', 4'-tri-0-p-Dglucopyranoside from leaves of Eruca

- sativa (Mill.). Phytochem., 57: 547-551.
- 21-Avila, J.(2014). Erucin, The major isothiocyanate in Arugula (Eruca Sativa) , inhibits proliferation of MCF7 tumor cells by suppressing microtubule dynamics.
- 22- El-Missiry, M.A. and El-Gindy, A.M. Choudhuri, S. (1999). Metallothionein: (2000). Amelioration of alloxan induced an intracellular protein to protect diabetes mellitus and oxidative stress in rats against cadmium toxicity. Annu Rev by oil of Eruca sativa seeds. Ann. Nutr. Pharmacol Toxicol., 39:267-294. Metab., 44: 97-100.
 - 23- Homady, M.H.; Hussain, H.H.; Tarawaneh, K.a. and Shakhanbeh, J.M. (2000). Effects of medicinal plant extracts used in Jordan on social aggression as well as testicular and preputial gland structure in male mice. J Bio Scien.,3(3):389-402.
 - 24- Morales, M. and Janick, J. (2002). Arugula: A promising specialty leaf vegetable. In: trends in new crops and new uses. Janick J, Whipkey A (Eds.). Alexandria, VA: ASHS Press, pp.: 418-423.
 - 25- Kothari, V.; Gupta, A.; Naraniwal, M.(2012).Comparative study various methods for extraction of antioxidant and antibacterial compounds from plant seeds. J Natural Remedies., Vol. 12(2): 162 - 173.
 - 26-Luna, L. G. (1968). Manual of histology staining, methods of armed forces. Institute of Pathology. 3rd edition. McGraw-Hill Book Company, New York and London.
 - 27-Ballesterose, R.; Bonsfills, N.; Chacon, N.; Garcia, J. and Gomez, E. (2012). Histomorphometery of the ligament using a generic-purpose image processing software: a new strategy for semi-automatized measurement. J Digit Imageing.,25: 527-536.
 - 28- Snedecor, G.W. and Cochran, W.G. (1973). Statistical Methods. 6th the Iowa state University press:238-248.

Sofikitis, N.; Giotitsas, N.: Tsounapi, P.; Baltogiannis, D. and Hormonal Pardalidis, N. (2008). regulation of spermatogenesis and spermiogenesis. J Steroid Biochem Mol Biol., 109:323-330.

No. (2)

- 30- Klaassen, C.D. Liu, J.
- 31- Nair, A.R.; DeGheselle ,O.; Smeets, K.; Van Kerkhove, E. and Cuypers, A.(2013). Cadmium-induced pathologies: Where is the oxidative balance lost (or Not). Int J Mol Sci., 14: 6116-6143.
- 32- Zakaria, A and Al-Busadah, K. (2015). Pentoxifyllin efficiency in protecting testes against cadmium toxicity . J Anim Vet Adv., 14 (1): 18-29.
- 33- Kamel, M.M.; Abd El Razek, A.H;. Ahmed, K.A. and Kamel, G.M. (2011). Exposure of adult male rats to cadmium: Assessment of sexual behavior, fertility, aggression as well as anxiety like behavior with special reference to biochemical and pathological alterations. Life J.,8(2): 106-119.
- 34-S.: Talaiekhozani, Alaee, A.;Rezaei, S. Alaee, K.: and Yousefian, E.(2014).Cadmium and male infertility. J Infert Reprod Biolo., 2(2): 62-69.
- 35- Miller, E. A.; Nudler, S. I.; Quinteros, F. A.; Cabilla, J. P.; Ronchetti, S. A.and Duvilanski, B. H. (2010).Cadmium induced-oxidative stress in pituitary gland is reversed by removing the contamination source. Hum Exp Toxicol., 29:873-880.
- 36- Jiménez-Ortega, V.; Cano Barquilla, P. Fernandez Mateos, P.; Cardinali, D. P. and Esquifino, A. I.(2012). Cadmium as an endocrine disruptor. Correlationwith anterior

aflatoxin b1 in male rabbits. Int J Acad

No. (2)

Res., 2: 67–74.

44- Koubaa, M.; Driss, D.; Bouaziz, F.; Ghorbel, R.E. and Chaabouni, S.E.(2015). Antioxidant and antimicrobial activities of solven extract obtained from rocket(Eruca sativa L.) flowers. Free Rad Antiox., 5(1):29-34.

45-Gunnarsson, D.; Nordberg, G.; Lundgren, P.and Selstam, G.(2003). Cadmium-induced decrement of the LH receptor expression and cAMP levels in the testis of rats. Toxicol. 183(1-3): 57-63.

46- Saeed, B.T.(2013). Effects of Cadmium on sperm parameters. histological and hormonal changes in testes of mature rats. Iraqi J Embryos and Infertility Researches., 3(6): 45-50.

47- Partra, R.C.; Rautray, A.K. and T.; Swarup, D. (2011). Oxidative stress in and amelioration. Vet Med Int., 2011:

Massanyi, M.; P.: M.; Omelka, Krajcovicova, V. and Duranova, A. Effects of subchronic exposure to cadmium and diazinon on testis and epididymis in rats. The Scientific World Journal. Volume 2014, Article 632581. pages. http://dx.doi.org/10.1155/2014/63258 1.

49-Al-Haddad, A.A.; Al-Okaily, B.N. and Hussein, S.M.(2008). Effect of different concentrations of cadmium chloride on some structural changes of testes in adult male rabbits. Basrah J Vet Res., 7 (2): 74-83.

pituitary redox and circadian clock mechanisms prevention and melatonin. Publicado en Free Radic Biol and Med ., 53: 2287-2297.

37- Elgawish, R.A. and Ghanem, M.E.(2014). Effect of long term cadmium chloride exposure testicular functions in male albino rats. Am J Anim Vet Sci., 9 (4): 182-

38- Ekpenyong, E.E.; Anderson, E.L.; Abdullateef, T.A.; Modupe, A.A.and Eunice, K.O. (2015). The impact of pudica on the mimosa histoarchitecture of hypothalamic pituitary testicular axis in cadmium rats . J Pharm Pharmaceutical Sci., 4 (10): 163-179. 39- Hamid, S.; Sahar, A.; Malik, F.; Hussain, S.; Mahmood ,R.; Ashfaq , K.: Malik. T.; Hassan, A. and Chaudhry, A.(2014).Physicochemical investigation and antioxidant activity studies on extracts of Eruca sativa seed., IJPC., 4(4):160-165.

- 40- Jin, J.; Koroleva, O.A.; Gibson, Swanston, J. and Magan, J. (2009). Analysis lead and cadmium toxicity and its of phytochemical composition chemoprotective capacity of rocket (Eruca 457327.doi: 10.4061/2011/457327. sativa and Diplotaxis tenuifolia) leafy salad 48-Adamkovicova, M.; Toman, R.; different Cabai, following cultivation in environments. J Agric Food Chem., 57(12): Martiniakova, 5227-5234.
 - 41- Nwofal, A. J.(2014). The role of ethanolic extract of salad rockets sativa) leaves on (Eruca the performance of male reproductive system in oxidative stressed rats. MSc. Thesis/College of Veterinary Medicine -University of Baghdad.
 - 42- Ansari, M.N.; Ganaie, M.A.; Khan , T.H. and Soliman , G.A. (2014) . Protective role of Eruca Sativa extract against testicular damage in streptozotocin-diabetic rats . IJBPAS, 3(7): 1067-1083.

50- Imran, A.; Muhammad, S. and Khalid, F.Y.(2003). Study of the effects of lead poisoning on the testes albino rats. Pak J Res.,42(3):97-101.

51- Aydos, K.; Guven, C.M. and Can, B. (2001). Nicotine toxicity to the ultrastructure of the testis in rats. BJU International 88, 622–626.

52- Wang, B.; Schneider, S.N.; Dragin, N.; Girijashanker, K.; Dalton, T.P.; He, L.; Miller, M.L.; Stringer, K.F.; Soleimani, M.; Richardson, D.D. and Nebert, D.W. (2007). Enhanced cadmium induced testicular necrosis and renal proximal tubule damage caused by gene-dose increase in a Slc 39a8- transgenic mouse line. Am J Physiol Cell Physiol., 292 (4): 523-535.

53- Jana, D.; Maiti, R. and Gosh, D. (2003).Effects of Stephania hernandifolia leafextract on testicular activity in rats. Asian J Androl., 5: 125-129.

54- Sinha-Hikim, I.; Artaza, J.; Woodhouse, L.; Gonzalez-Cadavid, N.; Singh, AB. and Lee, MI.(2002) Testosterone-induced increase muscle size in healthy young men is associated with muscle fiber hypertrophy. Am J Physiol. ,283:154-164.

55- Zitzmann, M. (2008). Effects of testosterone replacement and pharmacogenetics on physicalperformance and metabolism, Asian J. Androl., 10:364–372.

56-Nurzynska-Wierdak, R.(2015).Nutritional and energetic value of Eruca Sativa Mill . leaves . Acta Sci. Pol.,14 (4): 191-199.